

C. elegans Transcription Cofactors

Project Number: <BIO-EFR-0902>

<Bioinformatics Project for Walhout Lab Umass Medical School>

A Major Qualifying Project Report

Submitted to the Faculty

of the

WORCESTER POLYTECHNIC INSTITUTE

in partial fulfillment of the requirements for the

Degree of Bachelor of Science

in Biotechnology

by

Victor Zeng

Date: April 29th, 2010

Approved:

Prof. Elizabeth Ryder, Advisor

Table of Contents

TABLE OF CONTENTS	2
TABLE OF FIGURES	4
ABSTRACT	5
ACKNOWLEDGMENT	6
1 INTRODUCTION	7
2 BACKGROUND	9
2.1 TRANSCRIPTION	9
2.1.1 DNA	9
2.1.2 GENERAL TRANSCRIPTION FACTORS (GTFs)	11
2.1.3 REGULATORY TRANSCRIPTION FACTORS (TFs)	13
2.1.4 TRANSCRIPTION CO-FACTORS (TCFs)	14
2.2 OMICS	17
2.2.1 GENOME	17
2.2.2 OMIC DATABASES	18
2.2.3 INTERACTOME	19
3 METHODS	20
3.1 IDENTIFICATION OF TCF DOMAIN SEQUENCES	20
3.2 IDENTIFICATION OF ORTHOLOGOUS TCFs	21
3.3 EVALUATION OF THE PREDICTED TCF LIST	21
3.3.1 INTERACTOME DATABASES	21
3.3.2 INTERACTOME DATABASE PARSER	22
3.3.3 HIGH-THROUGHPUT YEAST-TWO HYBRID DATA	23
3.3.4 TCF TF CONVERGENCE	23
4 RESULTS	24
4.1 IDENTIFICATION OF PREDICTED TCFs	24
4.1.1 IDENTIFICATION OF HISTONE MODIFICATION PROTEINS VIA GENE CLASS SEARCHES	24
4.1.2 IDENTIFICATION OF HISTONE MODIFICATION PROTEIN VIA KNOWN DOMAINS	25
4.1.2 IDENTIFICATION OF NUCLEOSOME REMODELING COMPLEXES VIA TCF ORTHOLOGS	27
4.1.3 IDENTIFICATION OF HISTONE MODIFICATION INTERACTORS VIA KNOWN DOMAINS	27
4.1.4 DOMAIN BLAST RESULTS	28
4.1.5 IDENTIFICATION OF TCFs VIA ORTHOLOG SEARCHES	32
4.2 EVALUATING TCF PREDICTIONS WITH PHENOME DATA	32
4.3 EVALUATING TCF PREDICTIONS WITH INTERACTOME DATA	33
4.3 TCF TF OVERLAP	35
5 DISCUSSION	37
5.1 PREDICTED TCF DISTRIBUTION	37

5.2 PREDICTED TCF RNAi LETHALITY RATE	38
5.3 TCF TF CONVERGENCE	38
5.4 INTERACTOME EVALUATION	39
REFERENCES	40
APPENDIX A PREDICTED TCFS AND PHENOME DATA	44
APPENDIX B BLAST RESULT ALIGNMENTS	50
APPENDIX B.3 LSD-1 SPR-5 ALIGNMENT	50
APPENDIX B.2 BROMO-DOMAIN BLAST ALIGNMENT	50
APPENDIX B.3 CHROMO-DOMAIN BLAST ALIGNMENT	59
APPENDIX B.4 PLANT-HOMEO-DOMAIN BLAST ALIGNMENT	60
APPENDIX B.4 SET-DOMAIN BLAST ALIGNMENT	65
APPENDIX B.4 SET-DOMAIN BLAST ALIGNMENT	67

Table of Figures

FIGURE 1. NURF-1 TRANSCRIPT VARIANT	10
FIGURE 2. BASAL TRANSCRIPTION MACHINERY	12
FIGURE 3. TRANSCRIPTION REGULATORS	14
FIGURE 4. SET DOMAIN	26
FIGURE 5. JMJC DOMAIN	26
FIGURE 6. BROMO DOMAIN	27
FIGURE 7. CHROMO DOMAIN	27
FIGURE 8. PLANT HOMEODOMAIN	27
FIGURE 9. SET DOMAIN BLAST OUTPUT	28
FIGURE 10. JMJC DOMAIN BLAST OUTPUT	29
FIGURE 11. BROMO DOMAIN BLAST OUTPUT	30
FIGURE 12. CHROMO DOMAIN BLAST OUTPUT	30
FIGURE 13. PLANT HOMEODOMAIN BLAST OUTPUT	31
FIGURE 14. THE PREDICTED TCFs STATISTICS	33
FIGURE 15. TCF INTERACTOME MAPPING	34
FIGURE 16. INTERACTOME DATA ANALYSIS	35
FIGURE 17. TCF TF CONVERGENCE	36

Abstract

Transcription Cofactors (TCFs) are essential non-DNA binding gene expression regulatory proteins. 162 TCFs were predicted within *C. elegans* using literature search and BLAST. Predicted TCFs consist of mediators, TAFs, nucleosome remodeling, modification, and tail binding proteins. Using a proprietary PSI-MI2.5 parser, 98 known interactions were queried with only 9 interactions with predicted Transcription factors (TFs). 45.7% predicted TCFs shows cause embryonic lethality from RNAi phenotypes. The predicted TCFs can be experimented with predict TFs to find novel interactions.

Acknowledgment

There are many individuals that I would like to thank and acknowledge, as without their support this project would not have been possible.

I would like to thank Doctor Elizabeth Ryder, My WPI Advisor for her support and encouragement throughout my project.

Secondly, I would like to express my deepest appreciation to Doctor Marian Walhout, of the Umass Medical School for sponsoring my MQP. Her continual support and guidance has made this project a reality.

I would also like to express my gratitude to the members of the Walhout Lab for their invested interest and encouragement throughout the completion of my project.

1 Introduction

A biological system functions through the interactions of molecules such as carbohydrates, lipids, nucleic acids, and proteins. To fully understand the biological systems, biologists strive to complete the interactome, which contains all interactions of every molecule within an organism. Regulation of gene expression is an area of biological studies, and the number of molecules involved in the regulations and their interactions increase the intricacy of differential gene expression and dictate an organism's complexity (*Levine et al. 2003*). There are many protein components involved in the regulation of gene expression. General Transcription Factors (GTFs) are required for basal transcription and they are regulated by other factors in transcription. Regulatory Transcription Factors (TFs) are DNA binding regulatory factors that are not required for basal transcription. Transcription Co-Factors (TCFs) are non-DNA binding regulatory factors that are not required for basal transcription. GTFs, TFs, and TCFs are all essential for the regulation of transcription through their protein-protein interactions. A complex organism such as *Homo sapiens* has approximately 2600 predicted TFs (*Babu et al. 2004*). A simpler organism such as *Caenorhabditis elegans* (*C. elegans*) has a lower number of regulatory factors resulting in less molecular interactions, and causing it to be more feasible for systems biology studies than human. Studies of *C. elegans* molecular interactions within gene expression regulation will provide experiences and answers for future studies of *Homo sapiens*.

To understand the regulation of transcription, researchers must know which proteins are GTFs, TFs, and TCFs. Because of high level of conservation over evolution, the GTFs of *C. elegans* are indentified (*Verrijzer et al. 1995*). A DNA binding domain-based analysis generated a list of 934 *C. elegans* TFs (*Reece-Hoyes et al 2005*). There has been research with the effort in explaining TCF functions (*Roeder 2004*), but little is known of which proteins within *C. elegans* are TCFs. In addition, there is a lack of knowledge regarding the interaction between the three types of regulatory factors of transcription.

Currently, the *C. elegans* interactome is incomplete, and some of the recorded interactions are results of computational prediction that lacks experimental support. The

prediction of *C. elegans* TCFs will allow future protein-protein interaction detection with the predicted TFs and identified GTFs. The detection of these proteins' interaction can be done using high-throughput screening. The knowledge of their interaction will further the understanding of the transcription regulation network of *C. elegans*. In addition, the results of their interactions will aid the completion of the *C. elegans* interactome.

To accompany the development of the interactome, newer methods of data mining from the interactome database need to be developed. The size of the interactome database will increase with additional datasets causing searches within the interactome to be more difficult. Currently, there are multiple parties building interactome databases with different information. The current method permits the search of the interactions of only a single interactor within a database at a time. For biology studies using the interactome, biologists will need to gather all known interactions of many proteins in a time efficient manner. The creation of a program to perform batch interaction screening with all of the interactome databases will decrease the research time.

For this project, a list of predicted TCFs was determined for future detection of protein interactions, and a program was created for fast search of the interactome data. The prediction of TCFs required a comprehensive literature search for research regarding TCFs. Using the knowledge of the literature, protein families that relate to TCFs, and protein domains that show TCFs function were identified. The predicted TCFs were then gathered from the *C. elegans* genome through searches and Basic Local Alignment Search Tool (BLAST). Evaluation of the predicted TCFs then was done using the interactome data parser and the extensive *C. elegans* phenome.

2 Background

Gene expression regulations manage organisms' reproduction, development, and responses to external stimulus. Transcription, translation, localization, and degradation are some cellular processes where countless factors regulate organisms' gene expression. Transcription is the initial step in the central dogma, and it precedes the other cellular processes. The inhibition of factors that are involved in transcription regulation results in lethality (*Fraser et al. 2000*). This is because transcription regulation factors are key components of the gene expression regulation network. The mapping of transcription regulation factor interactions will improve the understanding of gene expression.

2.1 Transcription

Eukaryote transcription is a widely studied subject in biology due to its crucial role in the central dogma. Biologists view general transcription as a stepwise assembly line (*Dignam 1983*). General transcription consists of the initiation, elongation, and termination steps of RNA polymerization. In eukaryotes, the components of basal transcription are DNA, General Transcription Factors (GTFs), and RNA polymerases. Many different lineages of proteins that interact with the general transcription components emerged via evolution. There are additional proteins that interact with those proteins that interact with the general transcription components. Together all of the proteins produce a network of protein interactions that regulate transcription.

2.1.1 DNA

Transcription is the production of RNA using a template DNA. DNA contains multiple regions, and each region serves a critical role in the transcription process. In eukaryotes, the DNA has regions that are transcribed into RNA, promoter and enhancers that bind TFs, and many un-transcribed regions that form the tertiary structure of DNA. The interaction of proteins within these regions regulates the outcome of transcription.

The region of a gene that polymerases transcribe encodes the RNA transcripts. The transcribed RNA in eukaryotic organisms consists of sections termed exons and introns. The exons are selected and the introns are removed based on splice pattern to form the final transcript prior to translation (*Crick 1979*). This process produces transcription variants from one gene (Fig. 1). Sequences such as the start codon AUG and

the stop codons of UAA, UAG, and UGA can also be found within the RNA transcript. The Open Reading Frame (ORF) of the spliced mRNA that is located between the start and the stop codons will be translated. Based on the different splice variants, a different set of start and end codon may be encountered by the ribosome, and thus a different ORF is produced. Some of the transcribed fragments do not possess ORF, such as the DNA that encode enzymatic RNA. For those transcribed fragments that do possess ORFs, ribosomes can translate the resulting RNA transcripts into proteins (*Rosenberg et al. 1979*).

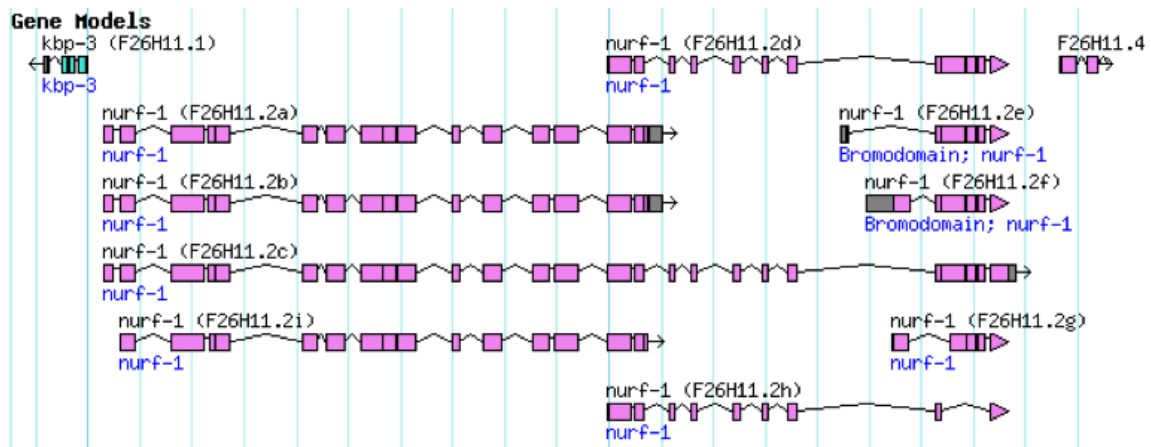


Figure 1. nurf-1 Transcript Variants

The 6 different transcript variants currently known for *C. elegans nurf-1* are shown.

RNA polymerases are initially recruited to the promoter and transcribe in a 5'→3' fashion, thus the promoter region of DNA is found at the 5' end of the transcribed region. The TATA box is a specific DNA sequence of TATAA within the promoter. The TATA Binding Protein (TBP) associates with the TATA box and creates a base for the assembly of the transcriptional machinery (*Nakajima et al. 1988*). Biologists refer to the combination of promoters and their ORFs as genes.

When genes are not activated for transcription, histones super coil DNA (*Almer et al. 1986*). Histones are chromatin structural proteins, and form nucleosome complexes with DNA (*Laybourn et al. 1991*). Nucleosome complexes are very compacted. This mechanism prevents most factors involved in transcription from accessing the DNA, and inhibits unsystematic transcription of the compacted genes. In contrast, the promoters of

activated genes are depleted of nucleosome, which allows the interaction between TFs and promoter sequence.

Enhancers are similar to the promoters because of their TF binding capability. Unlike the promoter, which must be located directly upstream of the regulated gene, the enhancer may be distal from the gene it regulates. In eukaryotes, the tertiary folding of DNA allows a distal enhancer to become extremely close to the gene it regulates. In some cases, the enhancer may exist on a completely separate chromosome as the regulated gene (Geyer *et al.* 1990). Finally, studies show some transcribed regions of DNA have TF binding affinity. For example, the murine immunoglobulin H μ core enhancer is located within the second intron of its regulated gene (Blackwood *et al.* 1998).

2.1.2 General Transcription Factors (GTFs)

Basal transcription in eukaryotes requires not only the DNA; it needs essential proteins that are termed GTFs, along with RNA polymerases. *In vitro*, GTFs are recruited to the promoter to form the Pre-Initiation Complex (PIC) with the RNA polymerases (Rowland *et al.* 1994). PIC is necessary for transcription because of its ability to recruit the RNA polymerases to genes being transcribed and aid the RNA polymerases with the down stream activity (Fig. 2).

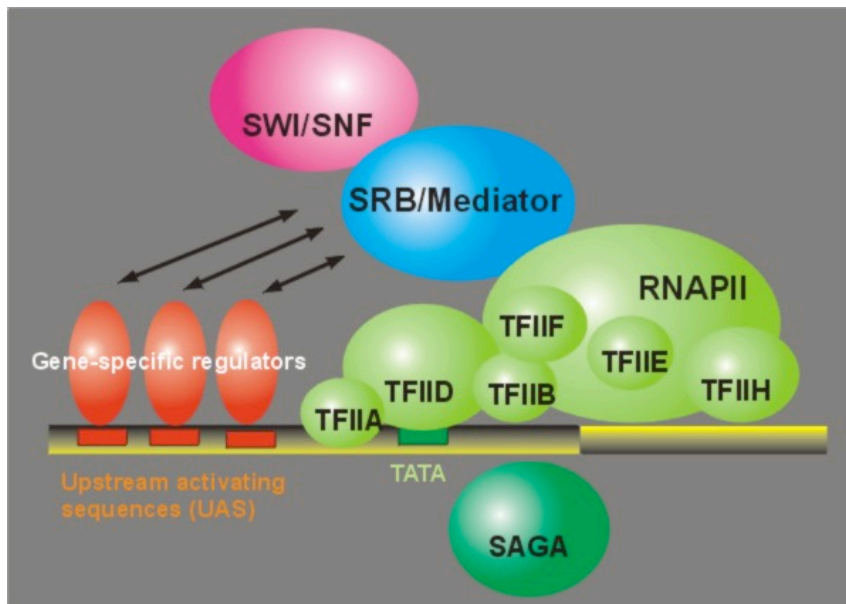


Figure 2. Basal Transcription Machinery

The basal transcription machinery is shown in light green, the mediator complex in blue, histone remodeling complexes in pink and dark green, and TFs in orange (Holstege *et al.* 1998).

One of the important roles of GTFs is the recruitment of the RNA polymerases. As described previously, the TATA box within the promoter is the base for the PIC for the transcription of mRNA via RNA polymerase II. TBP (TATA binding protein) is a subunit of GTF TFIID, and it is the base of a complex formed with TBP Associated Factors (TAFs) (*Lee. T. et al. 2000*). TAFs are distinguished from the GTF machinery in that they are not required for basal transcription. TFIIA could be considered either a GTF or a TAF, and it interacts with TBP in a similar manner as to TAFs. TFIIA is not required for basal transcription *in-vitro*, which initiated the debate of whether it is a GTF. TFIIA is essential *in-vivo* due to constitutive ubiquitous TFIID repressors in the nucleus (*Ozer et al. 1994*).

GTF TFIIB is another unit of the PIC that is required for basal transcription. TFIIB creates the bridge between TFIID and RNA polymerase II (*Verrijzer et al. 1995*). In addition, TFIIB is shown to have protein-protein interaction with TFs, such as the cAMP-response element binding protein (CREB) (*Tini et al. 2002*). TFIIB is regulated by CREB through protein-protein interactions, and these interactions are the key to understanding the regulation of transcription.

TFIIF binds DNA in a non-sequence specific manner, and is required for basal transcription in eukaryotes (*Robert et al. 1998*). TFIIF has protein affinity for both TFIIB and RNA polymerase II, and it is predicted to aid TFIIB in the bridging with RNA polymerase II. Though TFIIF has some structural functions in the PIC, its main purpose is to wrap DNA around the transcription complex. As the transcription complex travels down stream, DNA functions like a conveyer belt with the aid of TFIIF. Studies show phosphorylation of TFIIF may terminate transcription pauses (*Tan et al. 1995*).

Two GTFs are not involved in the recruitment of RNA polymerases to the transcription start site, but are still required for the PIC due to their duty during the elongation step of transcription. These two GTFs resemble TFIIF because phosphorylation of their c-terminal domain can also affect transcription pauses (*Kugel et al. 1998*). TFIIE is one of these GTFs, and its enzymatic function is DNA melting, which is to break the hydrogen bond of the double stranded DNA base pairing. TFIIE performs its activity through its zinc ribbon catalytic domain (*Okuda et al. 2004*). TFIIH functions

as a helicase in conjunction with TFIIE. TFIIH is identified as a helicase due to its ability to use cellular energy ATP to unwind the DNA helix and separate DNA during elongation. These GTFs are essential for granting the polymerase access to the DNA while moving down stream.

2.1.3 Regulatory Transcription Factors (TFs)

TFs are DNA-binding proteins that are not required for basal transcription. TFs have functions similar to the TBP in DNA binding and protein recruitment. TFs can both activate and repress transcription by recruiting or blocking the formation of PIC respectively. (Roeder 1996) Based on the DNA binding domain, the binding affinity of TFs may vary significantly. Some transcription factors have multiple DNA binding domains, which grants them more specificity for DNA interaction. There are many DNA binding domains for TFs. The majority of transcription factors have catalytic sites within their secondary structures. These secondary structures fit within the major groove of DNA allowing it to interact with the aromatic bases. (Mitchell *et al.* 1989) There are also TFs that bind to the minor groove of DNA, such as the TFs with the AT-hook domain. The AT hook domain does not bind a specific sequence but targets AT-rich regions of DNA. TFs typically bind to the enhancer and promoter regions of DNA. There have also been cases showing TFs association to heterochromatin (Raff *et al.* 1994). The DNA binding domains create an extensive network of interactions between TFs and DNA.

TFs also interact with bio-molecules other than DNA, such as proteins and lipids. The interactions with these bio-molecules regulate TFs binding with DNA. Heterodimerization and homodimerization of TFs create additional DNA specificity to the dimer, and affect DNA binding (Helin *et al.* 1993). The functions of some TFs are altered by different environments, such as hypoxia (Zheng *et al.* 1998). Nuclear hormone receptors are a group of TFs with ligands. These TF ligands interact with a variety of lipid hormones, such as estrogen, steroid, thyroid, vitamin A, and vitamin D receptors. These lipid hormones are hydrophobic and may penetrate the nuclear membrane for direct signaling to their target receptors (Evans 1988). TFs may also interact with non-TF proteins that result in different functions and regulations.

2.1.4 Transcription Co-Factors (TCFs)

TCFs are proteins that are involved in transcription that do not interact with DNA, and are not required for basal transcription. TCFs are typically recruited by TFs for their functions via protein-protein interactions (Roeder 2004). Most TCFs are found in complexes within the nucleus. TCFs are believed to have an assortment of distinct functions, including recruitment of transcription machinery, nucleosome remodeling, and histone modification (Fig. 3).

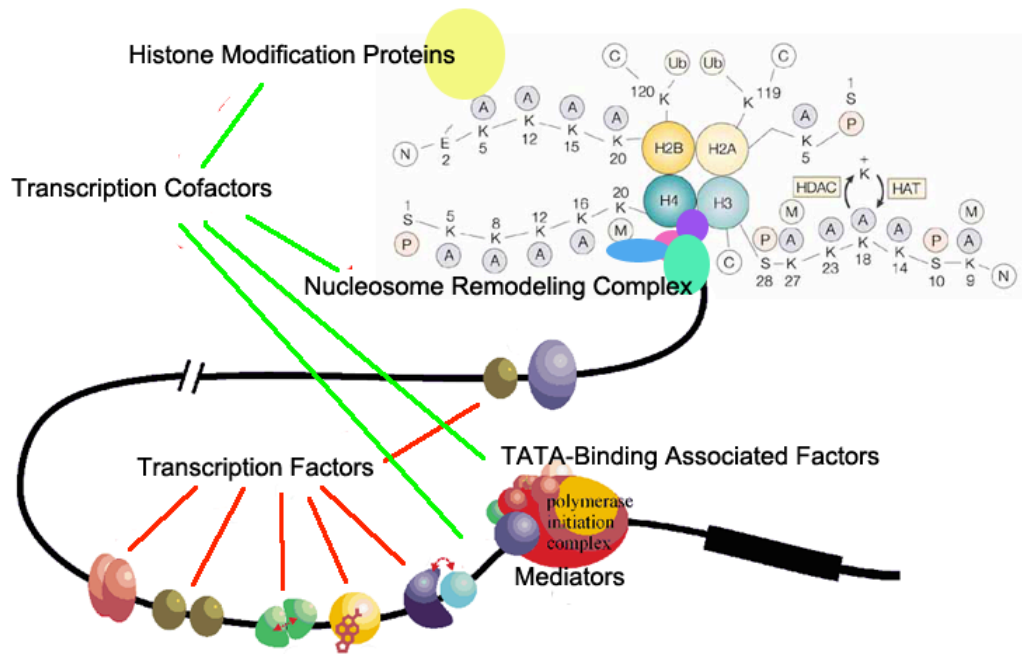


Figure 3. Transcription Regulators

The DNA is shown as a black line, and the protein-coding gene in a black box. The TCFs regulating this gene's expression in green lines, and the DNA bind TFs in red lines.

The Mediator Complex proteins are considered TCFs due to their ability to recruit GTFs to specific TFs. Mediator complex proteins resemble GTF TFIIB in ability to their bridge between TBP and polymerases. Mediator complex proteins vary based on the ligand of the TFs they bind. There are the ARC/DRIP mediators that interact with vitamin receptors, and the TRAP mediators that bind the thyroid receptors (Rachez *et al.* 2001). These mediators tend to associate with TFs activated by ligands to allow GTF and polymerase recruitment. They may also form complexes with other TCFs to recruit their enzymatic functions (Roeder 2004).

TAFs of GTF TFIID are considered TCFs. TAFs associate with TBP similar to TFIIA and TFIIB. TAF proteins can recruit and mask TBP from repressors and activators. Multiple TFs such as Sp1 require TAFs for bridging and recruitment of GTFs. (*Pugh et al. 1990*) TAFII250 also shows histone modification activities, and it highly resembles GCN5, a major yeast histone acetyl-transferase protein (*Mizzen et al. 1996*). TAFs can be considered as TCFs based on their ability to recruit transcription machinery and conduct histone modification.

Nucleosome remodeling is another function of TCF that work with histone modification to allow transcription machinery access to the naked DNA. There are many studied nucleosome-remodeling complexes such as the NuRD complex, Swi/Snf, RSC (*Roeder 2004*). These complexes all have very common features, such as proteins in the ATPase and helicase families. These nucleosome-remodeling proteins resemble TFIID in its function to modify DNA strand's conformation. DNA super-helices are flattened by nucleosome remodeling complex's helicases, while ATPases provide the kinetics for the physical movement (*Sudarsanam et al. 1999*).

Histone modification is the most important role of TCFs in the process of transcription. In eukaryotes, histone octamers cause the formation of heterochromatin from euchromatin, which inhibits transcription of the compacted region. The lysine rich histone tail has very basic chemical properties, and tightly binds to acidic DNA (*Allfrey 1964*). The lysines and arginines of histone tails are very susceptible to post-translational modification. Histone AcetylTransferases (HATs) work in pairs with Histone DeAcetylases (HDACs), and this reflects the high amount of changes in post-translation modification of histone that occurs in cells. ADP-ribosylation, methylation, phosphorylation, and summoylation are some other forms of histone modifications. These modifications create the histone code, which regulates transcription.

The major functions of TCFs are achieved through protein-protein interactions of either direct association, or post-translational modification. Using protein sequence consensuses, biology can predict possible protein domains for TCF activity. Some TCF domains were predicted based on their ability to interact with post-translationally modified proteins. The bromo-domain is a 110 amino acid peptide that folds to create

multiple alpha helices. It is a very important domain for some TCF due its ability to bind with acetylated lysine (*Zeng et al. 2002*). Many HAT and nucleosome remodeling proteins have the bromo-domain because it allows these proteins to localize to histone tails through recognition of histone acetylation.

The chromo-domain is 50 amino acids long and folds to create alpha helix and beta sheets that have specific protein affinity for methylated lysine. Peptide variability may cause chromo-domain proteins to associate with different methylated lysines of histone tails (*Cavalli et al. 1998; Brehm et al. 2004*). CHD-1 a highly conserved chromo-domain containing protein that binds to lysine-4 of the histone H3 tail, while the chromo-domain of Polycomb Protein binds to lysine-29 of the histone H3 tail. The chromo-domain of these two proteins only differs by 5 amino acids, thus, the selectivity may be caused by other factors. The chromo-domain's activity is essential for numerous TCF functions.

Plant Homeo Domain (PHD) finger is a cysteine rich protein domain that is approximately 50-80 amino acids long. This domain has distinct similarity to a zinc-finger, but does not have DNA binding capabilities. PHD activity is predicted to allow adhesion of protein complexes through direct association (*Aasland et al. 1995*). The PHD domain shows strong signs of self-association *in-vitro*, and it occurs in many proteins of chromatin remodeling complexes. The PHD domain also interacts specifically with trimethylated lysine.

Post-translational modification proteins exist in many other biological systems other than histones modification. Those proteins have high levels of similarity compare to the histone modification proteins, and cause the identification of a specific histone modification domain difficult. There are two domains known for their histone modification functions. The SET domain is a 130 amino acid peptide, and studies have connected it with transcription silencing and activation. The function of SET domain is methylation of histone lysine (*Dillon et al. 2005*). Different sub families of SET domain target different lysines of histone tails. SET domain is a major player in the histone code and is a TCF domain. Jmjc domain is the second post-translational modification domain,

and its functions as a histone methylase (*Klose et al. 2006*). Jmjc also contains multiple subfamilies that vary in peptide sequence.

2.2 Omics

Omics is a term used in biology that originated with the creation of the Genome and Proteome. Omics is the holistic approach in annotating all molecules of organisms. Currently, with the advancement of computer technology, omic information is annotated in computer databases. System biologists utilize engineering to develop new methods to streamline experimental process to provide the vast amount of data required for omic databases. These databases are stored in servers that allow biologists throughout the world to access the knowledge via the World Wide Web. Bioinformatics has emerged as a field for the analysis of the databases while making them easier access. There has been tremendous development in the storage, mining, visualization, and computation of omic databases.

2.2.1 Genome

As the oldest of the omic databases, the genome database is developed with sophistication. In 1989, Jean Thierry-Mieg of University of Montpellier and Richard Durbin of Sanger Institute developed “*A. C. elegans* DataBase” (ACeDB). ACeDB is an information system, and it is very different from traditional computer databases. (Biology Research Computer Hierarchy) Traditional filing in a database uses a family system, and this means the directory consists of parent, offspring, and siblings. In AceDB the file relationship is the user-defined, which is more suitable for storage of biological data. For example, user defined directory allows a hierarchy with gene, RNA transcript, ORF, and protein in order while allowing RNA transcript, ORF, and protein to be siblings under genes. Using user-defined directory can increase the mining speed in large biological databases.

An intelligent browsing system was also created to access the genome information generated from a variety of experimented data. Experts of bioinformatics created graphical browsers that allows biologists to navigate through the genome and conduct, data mining. www.wormbase.org is a browser of the AceDB. Using a graphical interface Wormbase can visually illustrate a specific locus of gene, whether

experimentally proven or predicted. This browser has *in-silico* abilities, which are computer predictions. Using different algorithms, Wormbase can determine signature features within the genome, such as AT-rich regions, and repeated sequences. In addition, the browser has an algorithm for Genome wide sequence alignment that allows biologists to search for particular sequence patterns. Other than the DNA sequence, the developers of the browser incorporated the ability to access information of other omic databases.

2.2.2 Omic Databases

There are many omic databases other than the genome. All of the omic databases work synergistically. The information of each database can be validated and cross-referenced by the others. Biological processes involve countless different molecules. During transcription, RNA polymerases transcribe template DNA into RNA. The transcriptome is an attempt to identify all RNA transcripts produced during transcription. A recent study of the *C. elegans* transcriptome has shown the 14% of sequenced transcripts do not have a corresponding gene in the genome. (*Shin et al. 2008*) This study of transcriptome has demonstrated missing information within the genome, and provided knowledge for future improvements.

During translation, the ribosome may translate RNA transcripts into protein. The ORFeome utilizes computation to predict possible ORFs using genome and transcriptome data (*Reboul et al. 2003*). The ORFeome prediction can also be validated using proteome data. The proteome is a collection of all proteins produced in an organism under all environmental conditions during all developmental stages. The verification of proteome data is easier with the creation of protein mass spectroscopy technique (*Mann et al. 1993*). With the knowledge of protein sequences, the ORF can be verified, and provide information for future studies involving transcription variants. This is another example of the synergy between omic databases.

Using the genomic information, system biologists built the phenome to study the phenotypic function of genes. The phenome incorporates phenotypic data for mutated genes' alleles as well from RNA Interference (RNAi) experiments. *In vivo*, micro RNA and small interfering RNA act by binding mRNA (*Fire et al. 1998*). The RNA Induced Silencing Complex (RISC) dice and break down the double stranded RNA to prevent

translation. Synthetic RNA can be created to perform RNAi on specific genes (*Kamath et al. 2002*). RNAi can be induced in organisms through multiple methods such as injection, feeding, and soaking during all different developmental stages (*Rual et al. 2004*). Using RNAi biologists may observe the phenotypic outcome resulting from the reduce expression of genes. Both experimental data of mutants and RNAi phenotype provide extensive information of gene functions.

Outside of the central dogma, many bio-molecules need to be included within omic database because of their involvement in gene expression. Carbohydrates and lipids are involved in organisms' metabolism, and can interact with proteins to produce differential gene expression. Carbohydrates such as glycans are involved in cellular signaling. The glycome is the annotation of carbohydrates in organisms. Lipids are also involved in cellular signaling. Molecules such as hormones and vitamins are annotated within the lipidome. The interactions of molecules within the glycome and the lipidome can be studied in conjunction with other omic databases to further the understanding of gene expression regulation.

2.2.3 Interactome

The Majority of Omics focuses on the annotation of bio-molecules. Interactome is developed to annotate the interactions between the bio-molecules of an organism. Studies predicted that human has approximately 650,000 protein interactions, which is about 3 times more than *C.elegans* (*Stumpf et al. 2008*). Numerous data are needed for the construction of an interactome. To perform the require number of repeated experiments, the aid of robotics automation and high-throughput system is used. High-throughput devices are designed to perform multiple experiments simultaneously. Micro array chips may be used to show more than hundreds of interaction at a time (*Bader et al. 2003*).

Many labs have been using high throughput yeast 2-hybrid system for gathering of large datasets to accomplish this goal. Yeast 2-hybrid system is able to determine binary protein interaction, and has been use by scientists for the mapping of *Saccharomyces cerevisiae* (*Ito et al. 1999*), *Drosophila melanogaster* (*Giot et al. 2003*), *C. elegans* (*Li et al. 2004*), and possibly in human (*Rual et al. 2005*). Protein mapping using interactome can provide a visualization of gene regulation networks.

3 Methods

The goal of this project was to create a list of *C. elegans* TCFs. There are many groups of TCF related proteins, such as histone modification and tail-binding proteins, nucleosome remodeling proteins, transcription mediators, and TAFs (Roeder 2004). Some proteins of these groups are incorporated into gene-classes that were gathered from Wormbase. Some proteins of these groups are identified as complexes, and were gathered through literature research. A few groups have conserved protein domains, and were gathered using BLAST of *C. elegans* genome.

3.1 Identification of TCF Domain Sequences

Based on countless previous researches, approximately 800 protein domains were identified through sequence alignment of various species' proteins with identical biochemical functions. Biologists use homologs, orthologs, and paralogs of domain containing proteins to determine the consensus domain sequences of functional peptide. The sequences of TCF protein domains were gathered from a database named Simple Modular Architecture Research Tool (SMART). SMART creates protein domain consensus sequences by aligning sequences of all proteins that are known to contain the specific domain. The protein sequences are gathered from databases, such as the National Center for Biotechnological Information (NCBI). SMART has low consensus accuracy for those protein domains that possess multiple sub families. This is because the functional peptides of those protein domains differ greatly between the sub families. The sequences of these domains are gathered through literature research.

BLAST was used to identify those proteins with the desired domain within the *C. elegans* genome. Those proteins with E-values less than e^{-10} were considered as a predicted TCF. Through literature research, the active sites and highly conserved amino acids of each protein domain of interest were gathered. Proteins with E-values higher than e^{-10} were kept as predicted TCF if all active sites and conserved regions matched within the alignment resulted from BLAST.

3.2 Identification of orthologous TCFs

Multiple TCF complexes were determined using previous research in multiple model organisms. The sequences of proteins that were identified as predicted TCF were used in a BLAST of *C. elegans* genome. To maintain the confidence of the orthologs gathered from BLAST, only results with E-value under e-50 were kept. Using this approach, many orthologs of the previously identified proteins were found. The majority of the orthologs found were already identified as predicted *C. elegans* TCFs through literature search. A literatures have predicted the orthologs of many non-*C. elegans* TCF complex proteins within *C. elegans*, they were also incorporated into the TCF list (Chue *et al.* 2006).

3.3 Evaluation of the predicted TCF list

The silencing of those proteins that are involved in the regulation of transcription can generate many different phenotypes. The phenotypes of gene silencing experiments were gathered from Wormbase to evaluate the specific lethality phenotype of predicted TCFs. Only the phenotypes from RNAi experiment were used for this analysis. Wormbase also stores the phenotypic outcome of many mutants. For the evaluation of the predicted TCFs, mutant phenotypes were not used. Although the level of gene silencing varies amongst genes, RNAi experiments are done with the same molecular approach. Based on the alleles of a gene, a range of different changes can occur to the gene expression. Certain mutations such as point mutation of a protein's active site may result in the complete loss of function. In other mutations, the protein's functions are not altered in the same manner, which may result in a range of different phenotypes. For the comparison of large groups of genes using phenome data, utilizing only RNAi phenotypes is more precise.

3.3.1 Interactome Databases

To better our understanding of gene expression, the interaction network of transcription regulation proteins needs to be completed. To determine the need for future experiments to be performed for the identification of novel interaction, predicted TCF data within the current interactome were evaluated. All of the interaction databases including Intact, MINT, and DIP are stored as plain ASCII text files using the Proteomics Standards Initiative – Molecular Interactions level 2.5 (PSI-MI 2.5) (Hermjakob 2006).

This file format adopts the XML structure that assigns classes to information using HTML tags, and subclasses are created within HTML tags of parent classes. The classes that can be assigned to data with PSI-MI 2.5 databases are dictated by the human proteome organization.

Currently, the *C. elegans* interactome is incomplete, and the majority of the data relies on orthologous interactions using the interactome data of other species. The interactome data are also spread amongst multiple databases, causing the search for interactions to be difficult. The European Molecular Biology Laboratories' European Bioinformatics Institute (EMBL-EBI) hosts the Intact protein interaction database for multiple major model organisms. The University of Rome Tor Vergata has created a database for the annotation of protein interaction termed Molecular INTeraction (MINT). Database of Interacting Protein (DIP) was created by the University of California Los Angeles. All of these databases have visual User Interfaces (UI) that allow users to search for interaction of a particular protein over the Internet. These UI lack the ability for batch protein interaction search that is required for the predicted TCFs. By creating a third party database parser, the tedious manual search can be avoided.

3.3.2 Interactome Database Parser

Perl is a widely used coding language that is heavily utilized in database servers. In addition, Perl is a powerful text parser, and can easily manipulate the text within PSI-MI 2.5. Perl was chosen as the language to program the interactome search software. The PSI-MI 2.5 also has a very intelligent method of separating interactome information. The interactors are stored within one section of the file, while the interactions and experimental information are stored separately in their own sections of the file. This method allows all information to be recorded once with the database, and the data parser only has to iterate through a single section of the file to gather specific data based on XML class tags. For the perl based search software, each piece of information within each section of the interactome is stored within an array. An associative array was then created by the program to link arrays of interactors, interactions, and experimental data together. By inserting a txt file of the gene names of interests, the software parses through the associate array to output known interaction information.

Using the perl PSI-MI 2.5 parser, the interactions of the predicted TCFs were gathered from both Intact and MINT. These interactions are shown in figure 4.4. DIP was not used because it did not match the same PSI-MI 2.5 standard as Intact and MINT. The Inconsistency of DIP XLM class compared to Intact and MINT caused error during the parsing of the perl search software, and the resulting information from DIP was invalid. In the future, the search software can be patched and debugged to allow for the usage of DIP data.

3.3.3 High-Throughput Yeast-Two Hybrid Data

The high-throughput yeast-two hybrid protein interaction detection data from the Vidal lab was also used to determine interactions of each predicted TCF (*Li et al. 2004*). The interactions detected from this experiment are not stored within a PSI-MI 2.5 file. Because the data is stored within an excel file, the perl search software could not be used. A basic perl parser was used to find interactions of the predicted TCFs within the Vidal lab data to prevent rigorous manual search. The interactions found using Vidal lab data increased the final number of interactions gathered for the predicted TCFs.

3.3.4 TCF TF Convergence

To identify the similarity between the predicted TCFs and previously predicted TF from Walhout lab, a comparison was done between the both lists. The result may provide further insight to the functionality of TCFs. The search of the convergence of the two lists was done using perl arrays. Information such as containing domains of those proteins that are blong to both lists was gathered from Uni-Prot.

4 Results

This project was conducted to create a predicted list of *C. elegans* TCFs (Appendix A). The prediction of *C. elegans* TCFs was done so future researchers may utilize the predicted proteins for the detection of interaction with other transcription related proteins, such as TFs, and GTFs. The prediction of *C. elegans* TCFs was carried out using data from previous TCF related research, such as the study of *C. elegans* TCF protein complexes, the study of TCFs of various eukaryotes, and the study of proteins with functions related to TCF functions. The predictions of *C. elegans* TCFs were analyzed using phenome and interactome data. In addition, to improve the mining of interactome data, a program was written to output interactions of the predicted proteins. This program was made to allow the search of multiple proteins' interactions using multiple interactome datasets at the same time.

4.1 Identification of Predicted TCFs

TCFs are non-DNA binding nuclear proteins that regulate cellular gene expression. Through literature research, TCFs were identified based on two functionalities. One is to regulate transcription through the histone code, and the other is to regulate transcription via the recruitment of transcriptional machinery. Both functionalities of TCFs involve proteins and complexes of different activities that can be categorized into sub-families of TCFs. The particular genes coding for each sub family within *C. elegans* were identified through the search of *C. elegans* gene classes, known domains, or TCF orthologs.

4.1.1 Identification of Histone Modification Proteins via Gene Class Searches

The histone code involves proteins with post-translational modification ability. There are many possible post-translational modifications of histones including acetylation, methylation, phosphorylation, sumoylation, and ADP-ribosylation. The particular gene coding for specific *C. elegans* histone modification proteins were identified through the search of *C. elegans* gene classes, and known domains. For acetylation, there is a particular *C. elegans* gene class of histone acetyl-transferase named *mys*, which were originally identified from histone acetylation complexes. The full name of *mys* is MYST, and it is the abbreviation of the 4 histone acetylation complexes, which

are MOZ, Ybf2/Sas3, Sas2 and Tip60. There are two *C. elegans* gene classes of histone deacetylase named hda and hdac. The proteins of these gene classes were gathered through Wormbase. These proteins were assigned to their gene class based on their public name. The public names of *C. elegans* genes are typically based on the major mutant or RNAi phenotype, but sometime the names are based on the predicted gene function.

There are also gene classes for those proteins involving the methylation of histones. Set is a gene class of histone methyltransferase, and it is name after the SET domain. A gene class of histone demethylase is named lsd, which stands for lysine specific histone demethylase. There is only one protein of this gene class in *C. elegans* that was discovered via its homology to the human lsd protein. One histone kinase family of *C. elegans* was found during the literature search. The air gene class in *C. elegans* is based on its homology to *Drosophila* Aurora kinase and yeast Ipl protein. These proteins were included as predicted TCFs.

Both summoylation and adp-ribosylation have literatures supporting their occurrences on histones (*Realini et al. 1992*). There was no literature showing specific ADP-ribosylase activity on histones within *C. elegans*. There are two gene classes of ADP-ribosylases within *C. elegans* genome, and they are arl and arf. The proteins of these two gene classes do not have experimental support of their activity on histones, and are not included as predicted TCFs. Ubc-9 was found to cause summoylation of histones within *C. elegans* (*R. Hay 2005*). Multiple proteins of ubc gene class are also ubiquitin-conjugating enzymes that are paralog of ubc-9. These proteins do not currently have literature supporting any histone activity, and are not included as predicted TCFs.

4.1.2 Identification of Histone Modification Proteins via Known Domains

Some protein domains are reported to have histone modification ability. The SET domain (*Dillon et al. 2005*), and Jmjc domain (*Klose et al. 2006*) were also chosen because of their unique histone methyltransferase and demethylase activities. The sequences of these two domains are shown below (Figs 4, 5). Set and Jmjc domain sequence were not gathered from SMART because they have multiple subfamilies, and did not have an accurate consensus sequence on SMART. Both sequences were gathered through literature along with the conserved amino acids and active sites.

ARSRIAGLGLYAKVDISMGDFIIEYKGEIIRSEVCEVREI 2420
 RYVAQNRGVYMFRIIDEWVIDATMAGGPARYINHSCDPNC 2460
 STQILDAGSGAREKKIITANRPIISANEELTYDYQFELEG 2500
 TTDKIPCLCGAPNCVKWMN

Figure 4. SET-Domain

The domain sequence of SET-domain gathered from set-16 is shown. The highlighted sequences are the lysine targeting amino acids (shown in yellow) and the catalytic site (shown in red) (Dillon et al. 2005).

(JHDM1)
 FSQTPLEDLVKSPQLVRQIDWVGQWPDALRQRWISFNDR 040
 DKKFYNPHTFPKQNYCLMSVANCYTDFHIDFSGTSVWY 080
 HVLKGRKVFVLIPTETNFFIYQEFIKTVNDNAFFGKSVE 120
 KCHVAILEPGDTMLIPSGWIHAVYTPDDSLVFGGNFLHSQ 160
 SCKTQLRVYQVEN

(PHF2/8)
 SDNNEMKEIAKPPRFVQEISMVNLWPDVSGAEYIKLLQR 040
 EEYLPEDQRPKVEQFCLAGMAGSYTDFHVDFFGGSSVYYHI 080
 LKGEKIFYIAAPTEQNFAAYQAHETSPDTTTWFGDIANGA 120
 VKRVVIKEGQTLIPAGWIHAVLTPVDSLDFGGNLFHLGN 160
 LEMQMRVYHL

(JARID1/2)
 GMCFSTFCWHTEDHWTYSVNYNHFGERKIWYGVGGEDA EK 040
 FEDALKKIAPGLTGRQRDLFHHMTTAANPHLLRSLGVPIH 080
 SVHQNAGEFVITFPRAYHAGFNEG

(JHDM3/JMJD2)
 DAQVEEWNMNLGTILEDTNIEIKGVNTVYLYFGMYKTTF 040
 PWHAEEDMDLYSINFLHFGAPKYWFAISSEHADRFRERFMSQ 080
 QFSYQNEYAPQCKAFLRHKTYLVTEPELLRQAGIPYATMVQ 120
 RPNEFIITFPRGYHMGFNLGYNLAESTNFASQRWIDYGKD 160
 AVLDCD

(UTX/UTY)
 KWGKQINELSKLPFCRLIAGSNMLSHLGHQVHGMNTVKL 040
 FMKVPGCRTPAHQDSNHMASININIGPGDCEWFAVPYEW 080
 GKMHLKCEKNGVDLLTGTFWPIIDLLDAGIPVHRFTQKA 120
 GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLL 160
 SYEY

Figure 5. Jmjc-Domain

The domain sequences of 5 sub-families of Jmjc-domain are shown. The highlighted sequences are the Fe (II) targeting amino acids (shown in yellow) and the α -ketoglutarate targeting amino acids (shown in red) (Klose et al. 2006).

There are many other forms of histone modification, and also many domains for acetyltransferase, kinase, ubiquitin conjugase, and adp-ribosylase. Those domains were not chosen as TCF domains because results from using a BLAST will produce many false positives. False positives are caused by proteins within *C. elegans* that possess those domains for post-translational modification and do not have direct activity on histones and the regulation of transcription.

4.1.2 Identification of Nucleosome Remodeling complexes via TCF Orthologs

The histone code modulates transcription through the recruitment of nucleosome remodeling complexes. Many nucleosome remodeling complexes have been discovered in model organisms, such as the SWR1/SRCAP of *A. thaliana*, the ISWI/NURF of *D. melanogaster*, the NuRD/CHD of *H. sapien*, and the SWI/SNF of *S. cerevisiae*. The *C. elegans* counterparts of these complexes were found through literature research. A study that used BLAST to determine the *C. elegans* orthology of each protein within each complex (Chue *et al.* 2006). TCFs found in this study were included in the predicted TCF list.

4.1.3 Identification of Histone Modification Interactors via Known Domains

There are protein domains with the ability to associate with the post-translational modification of histones discussed previously. The bromo-domain (Zeng *et al.* 2002), chromo-domain (Cavalli *et al.* 1998), and plant homeo-domain (Aasland *et al.* 1995) were chosen as TCF domains due to their specificity for binding to modified histone tails (Figs. 6,7,8). Proteins with these domains are prevalent in histone remodeling complexes, and other complexes that are involved in transcription regulation via the histone code.

```
PKRQTNQLQYLLRVVLTWVH----- 040
-----QFAWPFQQPVDAVKLNLPDYKIIKTP 080
MDMGTIKKRLNENY--WNAQECIQDFNTMFTNCYIYNK 120
-----PGDDIVLMAEALEKLFQKINEL 160
PT
```

Figure 6. Bromo-Domain

The domain sequence of bromo-domain was gathered from SMART. The highlighted sequences are done based on experiments of point mutations that cause loss of protein function (Zeng *et al.* 2001).

```
EYA-VEKIIDRR----- 040
-----VRKKGKVEY 080
YLKWKGYPETE-NTWEPENLD-----CQDLIQQYEAS 120
RK
```

Figure 7. Chromo-Domain

The domain sequence of chromo-domain gathered from SMART. The highlighted sequences are done based on the methyl recognition sites of the chromo-domain (Brehm *et al.* 2004).

```
FCR--VCKD----- 040
---GGELLCCD--TCP-SSYHI-HCLNPPLP----- 080
-----EI 120
PNGEWLCPRCT
```

Figure 8. Plant-Homeo-Domain

The domain sequence of Plant-Homeo-Domain gathered from SMART. The highlighted sequences are done based on the highly conservative cysteines residues (Aasland *et al.* 1995).

4.1.4 Domain Blast Results

A BLAST of *C.elegans* genome with the identified domain sequences (see Methods) was performed using Wormbase. The BLAST results for bromo-domain, chromo-domain, plant-homeo-domain, SET-domain and Jmjc-domain are shown in tables below. The E-value of these blast results are gathered and shown within the tables. In appendix B, the actual alignment of BLAST sequences are shown with the highlighted active sites and highly conserved amino acids.

Based on the e-value of set domain BLAST result, there were very high levels of alignments for all proteins (Fig. 9). There were also high levels of alignment of the catalytic site and lysine recognition site of SET-domain (shown in Appendix B.5). Many proteins of the set gene class were in the BLAST result, and were not included within this figure.

Gene-Name	Public Name	Blast E-Value
C43E11.3a	met-1	2e-19
C43E11.3b	met-1	2e-19
Y2H9A.1	mes-4	8e-12
R05D3.11	met-2	5e-10
R06A4.7	mes-2	1e-08
T12F5.4	lin-59	6e-08

Figure 9. Set-Domain BLAST Output

The BLAST results of SET-Domain sequence excluding those proteins of the set gene class with their alignment e-value are shown.

In general, BLAST with the five domain subfamilies of the Jmjc domain identified different proteins, with some overlap (Fig.10). PHF2/8 and JHDM1 identified identical proteins, and these are listed together in the table. All genes with E-values lower than the cut-off value of e^{-10} were included as predicted TCFs. Although psr-1 and T07C4.11 resulted in very high E-value from BLAST of JHDM1, the alignment has shown match for all of the important catalytic residues, and they were included as predicted TCF. Both rbr-2 and jmjd-2 were in the BLAST result of JARID1/2 and JHDM3/JMJD2 with low E-value, they were both included in the table once. The E-values of all genes other than rbr-2 and jmjd-2 resulted from the BLAST JARID1/2 and JHDM3/JMJD2 were high, the majority of them had alignment of the α -ketoglutarate targeting residue, but not the Fe (II) targeting residue (shown in Appendix B.6). These

proteins were not included as TCFs. tag-279 and C29F7.6 did result with low E-value during the BLAST of UTX/UTY, and matching on each catalytic residues (shown in Appendix B.6). tag-279 and C29F7.6 were included as TCFs.

Gene-Name	Public Name	Blast E-Value
JHDM1 and PHF2/8 subfamilies		
T26A5.5a	T26A5.5	e-108
T26A5.5b	T26A5.5	e-106
F29B9.2a	F29B9.2	2e-39
F29B9.2b	F29B9.2	2e-39
F43G6.6	F43G6.6	6e-32
F29B9.4a	psr-1	4e-05
T07C4.11	T07C4.11	5e-05
F29B9.4b	psr-1	5e-05
JARID1/2 subfamilies		
ZK593.4	rbr-2	5e-65
Y48B6A.11	jmjd-2	2e-18
C29F7.6	C29F7.6	0.14
C16C10.2	C16C10.2	0.73 *
F23D12.5	F23D12.5	0.91 *
JHDM3/JMJD2 subfamilies		
C29F7.6	C29F7.6	9e-07
F18E9.5b	tag-279	5e-04
F18E9.5a	tag-279	5e-04
F23D12.5	F23D12.5	0.031
UTX/UTY subfamilies		
D2021.1	utx-1	e-105
F18E9.5b	tag-279	6e-43
F18E9.5a	tag-279	1e-35
C29F7.6	C29F7.6	1e-31

Figure 10. Jmjc-Domain BLAST Output

The BLAST results of 5 different sub families for jmjc-Domain with their alignment e-value are shown. Genes shown with an asterisk in its BLAST e-value were not included as predicted TCF.

The e-values of many bromo-domain BLAST results were high (Fig. 11). There were high levels of alignment of the sites that caused the loss of function via mutation, thus most of these proteins were included in the TCF list (shown in Appendix B.2). The EGF receptor received a very high BLAST E-value, which demonstrates the lack of alignment. In addition, only one of the 5 loss of function mutation sites is aligned for EGF receptor, and was not included as an TCF. The transcript variants of each gene received identical E-values and alignment except for the h transcript variant of nurf-1. The alignment of this variant of nurf-1 showed only 2 of the 5 loss of function mutation sites matched with the bromo-domain sequence.

Gene-Name	Public Name	Blast E-Value
F57C7.1a	Female Sterile Homeotic Protein	8e-30
F57C7.1b	Female Sterile Homeotic Protein	2e-28
Y119C1B.8a	tag-332	7e-27
Y119C1B.8b	tag-332	1e-26
F13C5.2	Bromodomain Containing Protein	8e-16
H20J04.2	H20J04.2	2e-13
R10E11.1c	cbp-1	4e-13
R10E11.1b	cbp-1	4e-13
R10E11.1a	cbp-1	4e-13
F26H11.2e	nurf-1	2e-12
F26H11.2f	nurf-1	2e-12
F26H11.2d	nurf-1	2e-12
F26H11.2g	nurf-1	2e-12
F26H11.2c	nurf-1	3e-12
Y47G6A.6	pcaf-1	2e-11
C26C6.1a	pbrm-1	4e-09
F01G4.1	psa-4	5e-08
ZK783.4	flt-1	3e-07
C01H6.7a	tag-298	9e-07
C01H6.7b	tag-298	1e-06
W04A8.7	taf-1	1e-05
F11A10.1c	lex-1	1e-04
F11A10.1b	lex-1	2e-04
F11A10.1a	lex-1	2e-04
F26H11.2h	nurf-1	0.20
C34C6.3	EGF receptor	0.40 *

Figure 11. Bromo-Domain BLAST Output

The BLAST results of Bromo-Domain with their alignment e-value are shown. Genes shown with an asterisk in its BLAST e-value were not included as predicted TCF.

The E-values of all chromo-domain BLAST results were high (Fig. 12). The high E-value resulted because chromo-domain has multiple sub-families, and the domain consensus of all sub-families gathered from SMART is different from the chromo-domain sequence found in *C. elegans*. There were high levels of congruence of the methyl recognition sites during alignment (shown in Appendix B.3), and all of the BLAST results of chromo-domain were included as TCFs.

Gene-Name	Public Name	Blast E-Value
K08H2.6	hpl-1	2e-05
ZK1236.2	cec-1	0.010
K01G5.2c	hpl-2	0.11
K01G5.2b	hpl-2	0.11
F32E10.2	Chromo-domain Containing Protein	0.24
K01G5.2a	hpl-2	0.53

Figure 12 . Chromo-Domain BLAST Output

The BLAST results of Chromo-Domain with their alignment e-value are shown.

The E-values of many plant-homeo-domain BLAST results were high (Fig. 14). Though the E-values were high, there were high levels of alignment of the highly conserved cysteine residues of Plant-Homeo-Domain (shown in Appendix B.4). All PHD BLAST results were included as predicted TCFs.

Gene-Name	Public Name	Blast E-Value
T13G8.1	chd-3	3e-17
F26F12.7	let-418	4e-12
ZK783.4	flt-1	2e-09
C44B9.4	athp-1	2e-07
T12D8.1	set-16	2e-07
ZK593.4	rbr-2	4e-07
F17A2.3	PHD-finger Protein	6e-06
Y59A8A.2	Y59A8A.2	2e-04
K09A11.5	PHD-finger Protein	2e-04
C28H8.9a	C28H8.9a	5e-04
F33E11.6b	F33E11.6b	0.003
H05L14.2	Zinc finger C3HC4 type Protein	0.010
F26H11.2i	nurf-1	0.023
F26H11.2b	nurf-1	0.023
F26H11.2a	nurf-1	0.023
F26H11.2c	nurf-1	0.027
H20J04.2	H20J04.2	0.049
F42A9.2	lin-49	0.051
C11G6.3	PHD-finger Protein	0.083
F54F2.2a	zfp-1	0.20
Y51H1A.4	ing-3	0.20

Figure 13. Plant-Homeo-Domain BLAST Output

The BLAST results of Plant-Homeo-Domain with their alignment e-value are shown.

4.1.4 Identification of PIC Recruitment TCFs via Gene Class Searches

Multiple *C.elegans* TCF complexes were determined based on previous research. These TCF complexes include mediator complexes, such as Activator Recruited Complex (ARC), Cofactor Required for SP1 (CRSP), and Thyroid Hormone Associated Proteins (TRAP) (Rachez *et al.* 2001). These mediator complexes bind TF to recruit RNA polymerase. Previously, all of the *C. elegans* proteins within these mediator complexes were renamed with the mdt prefix for their public name (Bourbon *et al.* 2004). These proteins can be gathered from Wormbase using a global search of the mdt gene class. There were 6 proteins of mdt prefix that are not included within the mdt gene class due to their previous public names. These 6 proteins were manually gathered from Wormbase. The *C.elegans* TAF complex has also been previously addressed as a TCF complex

(Roeder 2004). The proteins within the TAF complexes were gathered from WormBase through search of the taf gene class.

4.1.5 Identification of TCFs via Ortholog Searches

Using BLAST, many orthologs of TCFs were identified. All of the TCFs but one identified in this manner were previously identified through other methods, such as domain based search, and literature research. The one gene that was new to the TCF list was spr-5. This gene is an ortholog of lsd-1, a histone methyltransferase. spr-5 had a very high alignment with lsd-1 with an e-value of e^{-147} . spr-5 is very likely a paralog of lsd-1.

4.2 Evaluating TCF Predictions with Phenome Data

Due to the heavy involvement of TCFs in the gene expression regulation network, the gene silencing is predicted to produce a high lethality rate. The phenome was accessed specifically for searches of phenotype related to lethality, which are characteristics of worms that die prematurely during any stage of the life cycle. Based on a large scale RNAi experiment of *C. elegans* chromosome I 5.5% of the genes result in embryonic lethality (Kamath et al. 2003). A high percentage of embryonic lethality phenotype shown the genes is related to transcription regulation, and more likely to be TCFs. This evaluation of RNAi lethality is a low estimate of 46% embryonic lethal, because those genes without any RNAi experimental data in the phenome are considered as negative for RNAi lethality. The RNAi phenotypes gathered are shown in Appendix A.

Based on the data gathered from the phenome of each sub group of TCFs, methyltransferase have the highest distribution of the overall list, and have the lowest lethality rate. Corresponding, the histone methyl-binding domains also have a low lethality rate. The chromatin remodeling complexes all had a very high lethality rate (Fig. 14).

Categories	Count	Emb Lethal	Larval Lethal	Lethal	General Lethality	Percentage
Total	162	74 (46%)	32 (20%)	33 (20%)	86 (53%)	100%
Complex Proteins	73	45 (62%)	21 (29%)	20 (27%)	53 (73%)	45%
TAF	17	8 (47%)	1 (5.9%)	4 (24%)	9 (53%)	10%
Mediator	23	17 (74%)	6 (26%)	5 (22%)	20 (87%)	14%
SWI/SNF	10	5 (50%)	6 (60%)	5 (50%)	9 (90%)	6.2%
NuRD/CHD	3	2 (67%)	2 (67%)	2 (67%)	3 (100%)	1.9%
ISWI/NURF	4	3 (75%)	2 (50%)	2 (50%)	3 (75%)	2.5%
SWR1/SRCAP	7	5 (71%)	1 (14%)	2 (29%)	5 (71%)	4.3%
COMPASS	3	1 (33%)	1 (33%)	0 (0%)	1 (33%)	1.9%
NuA3	1	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0.6%
TIP60/NuA4	4	3 (75%)	2 (50%)	0 (0%)	3 (75%)	2.5%
Histone Modification	60	19 (32%)	7 (12%)	10 (17%)	22 (37%)	37%
Acetyltransferase	6	3 (50%)	1 (17%)	1 (17%)	4 (67%)	2.5%
Deacetylase	7	3 (43%)	2 (29%)	1 (14%)	3 (43%)	4.3%
Methyltransferase	35	7 (20%)	3 (8.6%)	3 (8.6%)	9 (26%)	22%
Demethylase	11	4 (36%)	1 (9.1%)	4 (36%)	5 (45%)	6.8%
Kinase	2	2 (100%)	0 (0%)	1 (50%)	2 (100%)	1.2%
Summoylase	1	1 (100%)	0 (0%)	1 (100%)	1 (100%)	0.61%
Histone Tail Binding	32	14 (44%)	4 (13%)	4 (13%)	15 (47%)	20%
Bromo-domain	13	8 (62%)	3 (23%)	2 (15%)	9 (69%)	8.0%
Chromo-domain	4	1 (25%)	0 (0%)	0 (0%)	1 (25%)	2.5%
PHD	15	5 (33%)	1 (6.7%)	2 (13%)	5 (33%)	9.3%

Figure 14. The Predicted TCF List Statistics

The numbers and percentages of each type of TCF found in the current predicted list of TCF are shown. In addition, this table shows the number and percentage of the genes within the predicted TCF list that demonstrates the selected phenotypes of lethality using RNAi data. Emb lethal means the worm dies in the embryonic stage, larval lethal means the worm dies in the larval stage, and lethal means the worm dies prematurely not in a developmental stage. General lethality measures whether a gene have any of the three described phenotypes. Those sub families of TCFs shown in red have a lower percent of lethality rate compare to others.

4.3 Evaluating TCF Predictions with Interactome Data

A goal of this project was to determine the number of known interactions exist for the predicted TCFs. A perl based computer software was created to automatically search through multiple databases for known interactions quickly. Overall, 98 interactions were found within all the interactome databases queried with the yeast-2-hybrid data provided by the Vidal lab (Fig. 15). Only 9 of those interactions were between TCFs and TFs. Based on the theories of TCFs and TFs functionalities, many more interactions are currently unknown.

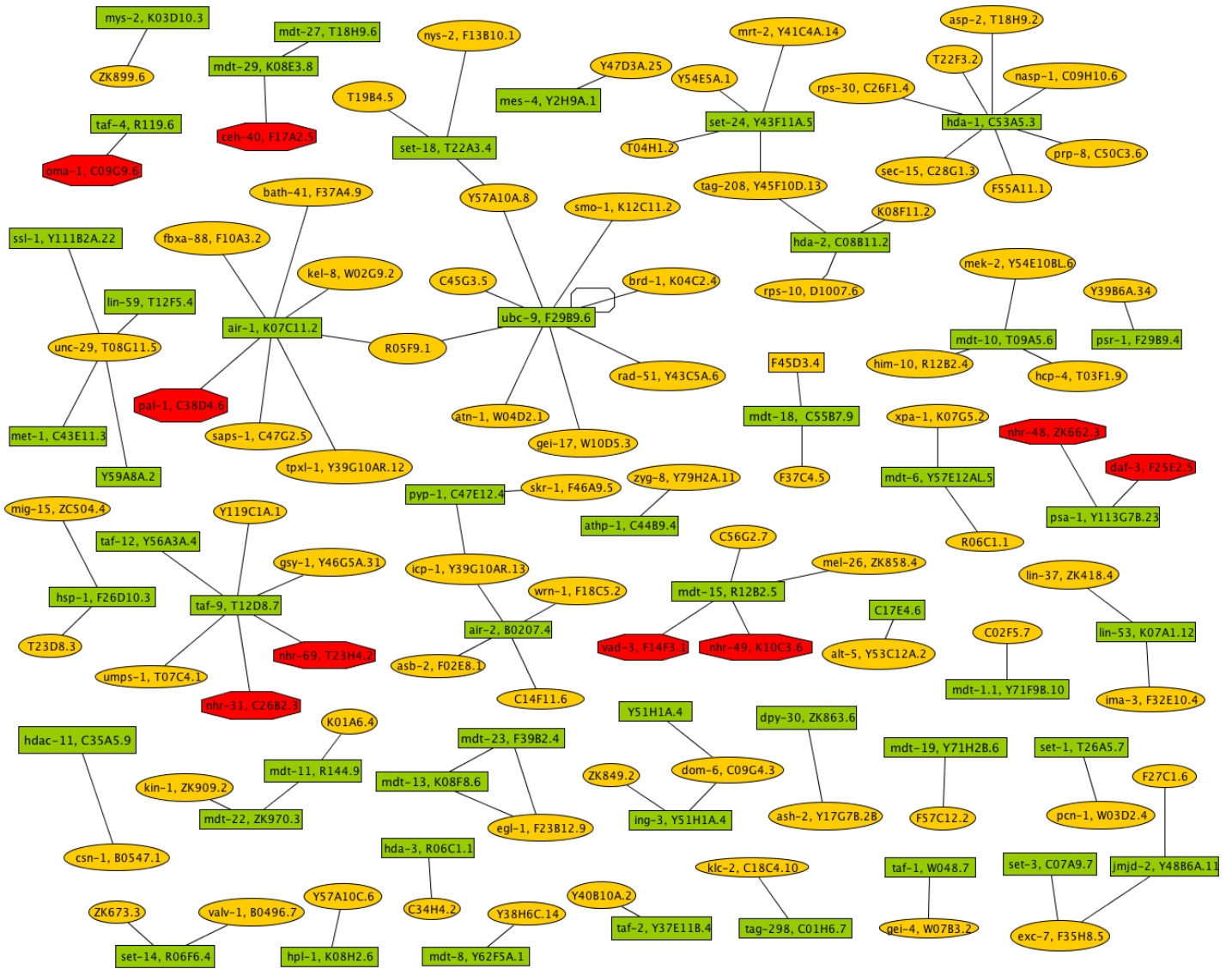


Figure 15. TCF Interactome Mapping

The known interactions of the predicted TCFs shown in green, the predicted TFs shown in red, and non-TCF-TF proteins shown in yellow.

Based on the predicted number of interactions within *C. elegans* and the current number of proteins within the proteome, there are approximately 4.8 interactions per protein (Fig. 16). Because of the functionality of TCFs, more interactions are expected, compared to other cellular proteins, which makes the 4.8 a low estimate of interactions

per TCF. The data from this project has shown 98 interactions of TCFs, between the lists of 162 predicted TCFs, there are only 0.6 interactions per protein. The interactome is far from complete because 0.6 is much lower than the already low estimate of 4.8 interactions per protein.

Categories	Interactors	%	Interactions	%	Interactions per Interactors
Overall	24,202	100%	116,000	100%	4.8
Intact	2,854	11.8%	3,520	3.0%	1.2
MINT	3,678	15.2%	3,503	3.0%	1.0
Vidal Lab	2,608	10.8%	8,378	7.2%	3.2
Predicted TCFs	162	0.7%	98	<0.1%	0.6

Figure 17. Interactome Data Analysis

The number of overall protein interactors within *C.elegans* according to Sanger Institute's proteome is shown. The number of interactions is based on the predicted size of *C.elegans* interactome (Simonis et al. 2009). The number of interactors and interactions of Intact and MINT are determined using the interaction detection software with the available data. The number of interactors and interactions of Vidal Lab data are calculated from the provided spreadsheet. This table also shows the number of interactors and interactions within the interactome map of the predicted TCFs, and the number of TCFs that are interactors within the map.

4.3 TCF TF Overlap

The final list of predicted TCFs were compare to the predicted TF list created from a previous project. Few genes were used in both lists (Fig. 17). The predicted TF list was created using all proteins possessing a DNA binding domain, and shown to bind DNA through experimentation. TCFs are believed to be non-DNA binding gene expression regulators, so those proteins that are also within the predicted TF list have a possibility of being non-TCFs.

Gene-Name	Public Name	TF Feature	TCF Feature
F15E6.1	set-9	AT Hook	Histone Methyltransferase
C01G8.9	let-526	ARID/BRIGHT	SWI/SNF Complex
Y113G7B.23	psa-1	MYB	SWI/SNF Complex
Y71H2AM.17	Y71H2AM.17	HMG Box	SWI/SNF Complex
F37A4.8	isw-1	AT Hook, MYB	ISWI/NURF Complex
C17E4.6	C17E4.6	YL1 TF	SWR/SRCAP Complex
Y105E8A.17	ekl-4	MYB	SWR/SRCAP Complex

Figure 17. TCF TF Convergence

A group of genes that were predicted as both TCFs and TFs is shown. This figure as shows the DNA binding domain that resulted these genes to be predicted as TFs. In addition, the functional features that resulted in the TCF prediction are shown in the figure.

From looking closely at the DNA-binding domains possessed by each gene shown to be in both predicted TCF and TF lists, it was determined that they were all non-specific DNA-binding domain except for C17E4.6. C17E4.6 was gathered for the predicted TF list base on a literature that provided experimental result of DNA-binding, and no DNA specificity was specified.

5 Discussion

Through this project a list of 162 *C. elegans* proteins were predicted as TCFs. The predicted list consists of a variety of different proteins. The RNAi phenotype and distribution of each predicted TCF were analyzed. The interactome was also evaluated for possibility of future expansion. Through these analyses, many new hypotheses were made that can be tested in future research of gene expression.

5.1 Predicted TCF Distribution

To achieve a high level of confidence with the predicted TCFs, only those proteins that have previously been studied as TCF, or possess known domains that are related to TCF functions were gathered. Proteins with histone methylation function had the biggest representation within the predicted TCF list, with 35 predicted methyltransferases and 11 predicted methylases. These proteins comprise 28.4% of the predicted TCF list. Other histone modification proteins were fewer in numbers, with the most common being proteins with histone acetylation function that encompass 6.8% of the predicted TCF list. This difference may be due to the number of previous studies conducted on the different types of histone modification proteins.

The majority of methylation activity within eukaryotic cells is DNA methylation and histone methylation. Based on previous studies, there are no signs of DNA methylation within *C. elegans*, thus explaining the lack of DNA methylation proteins (Bird 2002). This finding simplified the characterization of histone methylation proteins within *C. elegans*. In contrast, other proteins that perform post-translational modification on histones (e.g. kinases, acetylases) have many homologs that have identical enzymatic functions. For example, the kinase domain of *air-1* has more than 50 proteins that have lower than e^{-25} alignment using Wormbase BLAST. None of those proteins other than the two Aurora kinases has literature supporting any histone modification activity. It is possible that many *air-1* homologs may modify histone proteins, but not enough experimental data are available to know; thus, none of these proteins except the Aurora kinases were included in the predicted TCF list. This complication in the histone modification protein identification process may have created the difference between the numbers of each type of histone modification proteins gathered within the TCF list.

Although certain difficulty exists during the identification of specific histone modification proteins, there are still hypotheses that can be made based on the findings within this project. There is a potential that histone methylation is the primary method of transcription regulation via histone code within *C. elegans*. However, equal numbers of proteins were found that binds histone tails modified by methylation and acetylation, which does not support the hypothesis. Future experiments can be done on all the *C. elegans* summoylase, adp-ribosylase, kinase, and acetylase to determine histone activity.

5.2 Predicted TCF RNAi Lethality Rate

There were many gene-silencing experiments performed previously on the predicted TCF proteins, and they are annotated within online databases. The RNAi data from these experiments were used for the analysis of the predicted TCFs. RNAi data comparison of each predicted TCF group showed that histone methylation proteins have a lower percentage of lethality compare to other types of histone modification. The RNAi data of the histone tail-binding domain matches the data of histone modification proteins. Histone tail lysine-acetylation binding bromo-domain proteins have a higher lethality phenotype percentage than lysine-methylation binding proteins of chromo-domain and PHD. These data suggest that methylation may regulate less essential pathways.

Overall, the RNAi phenotypes of all 162 genes showed that 46% of them are embryonic lethal and 53% of them have showed some form of lethality. Comparison to the result of 5% embryonic lethality from the genome wide RNAi of chromosome I (*Kamath et al. 2003*) shows this group of 162 genes have distinct characteristics, and are not a random selection. TCFs are believed to be centralized in the regulation net work, and their silencing may result in the silencing of the production of many other proteins. The high lethality rate of RNAi provides evidence that the predicted TCFs have regulatory functions on general transcription, and the expression of other proteins.

5.3 TCF TF Convergence

The comparison between the predicted TCFs and TFs has shown some duplication between the two lists. There are in total 7 proteins that exist in both lists, and each of these 7 proteins possesses DNA binding domains that caused them to be predicted as TFs. The domains of all 7 proteins are non-specific DNA binding domains,

such as high mobility group, MYB. 6 out of the 7 proteins are a part of the predicted TCF complexes. It is possible that the non-specific DNA binding domains are utilized for the localization of TCF complexes. Experiments can be done on those predicted TCF proteins with DNA binding domain. Point mutation can be performed on the DNA binding domain to observe the outcome. If the proteins are non-active to perform their function due to the site mutation, then the non-specific DNA binding domain can be used as a method to determine more TCFs.

5.4 Interactome Evaluation

TCFs are a vital component of eukaryotic transcription regulation. There are 24,202 proteins currently recorded in the *C. elegans* proteome. Approximately 900 to 1500 of those proteins are predicted as TFs. Using the current interactome data, only 98 interactions are found with the predicted TCFs, and only 9 interactions with the predicted TFs. TCFs are theoretically predicted to be highly interactive with TFs.

The predicted completed interactome using the current proteome count and full interaction prediction estimates 4.8 interactions per protein. Using the current interactome data, only 0.6 interactions per protein were found for the highly interactive TCFs. These data illustrate that the current interactome is far from complete. The determination of novel interaction for the interactome will further our understanding of the transcription regulation. A high throughput yeast 2 hybrid screen of all predicted TCFs and TFs will likely yield a large number of novel interactions.

References

- Aasland R., Gibson T. J., and Steward F. A., "The PHD finger: implication for chromatin-mediated transcriptional regulation". *Elsevier Science* 4.965 (1995): 56-59.
- Allfrey V. G., Faulkner R., and Mirsky A. E., "Acetylation and methylation of histones and their possible role in the regulation of RNA synthesis." *PNAS* 51.5 (1964): 786-94.
- Almer A., Hans R., Albert H., Wolfram H., "Removal of positioned nucleosomes from the yeast PHO5 promoter upon PHO5 induction releases additional upstream activating DNA elements". *EMBO J* 5 (1986): 2689-96.
- Babu M. M., Luscombe N. M., Aravind L., Gerstein M., Teichmann S. A., "Structure and evolution of transcriptional regulatory networks". *Curr. Opin. Struct. Biol.* 14.3 (2004): 283-91.
- Blackwood E. M., and Kadonaga J. T., "Going the distance: a current view of enhancer action". *Science* 281 (1998): 60-3.
- Brehm A., Tufteland K. R., Aasland R., and Becker P. B., "The many colours of chromodomains". *BioEssays* 26 (2004):133-40.
- Cavalli G., and Paro R., "Chromo-domain proteins: linking chromatin structure to epigenetic regulation". *Current Opinion in Cell Biology* 10.3 (1998): 554-60.
- Crick F. "Split genes and RNA splicing". *Science* 204.20 (1979): 263-71. Dignam J. D., Lebovitz R. M., and Roeder R. G., "Accurate transcription initiation by RNA polymerase II in a soluble extract from isolated mammalian nuclei". *Nucleic Acids Research* 11.5 (1983): 1475-89.
- Evans R. M., "The steroid and thyroid hormone receptor superfamily". *Science* 240.4854 (1988): 889-95.
- Fire A., Xu S., Montgomery M. K., "RNA as a target of double-stranded RNA-mediated genetic interference in *Caenorhabditis elegans*". *PNAS* 95 (1998) 15501-7.
- Fraser A. G., Kamath R. S., Zipperlen P., Martinez-Campos M., Sohrmann M., and Ahringer J., "Functional genomic analysis of *C. elegans* chromosome I by systematic RNA interference". *Nature*. 408 (2000): 325-30.
- Geyer P. K., Green M. M., and Corces V. G., "Tissue-specific transcriptional enhancer may act in trans on the gene located in the homologous chromosome: the molecular basis of tranvection in *Drosophila*". *EMBO* 9.7 (1990): 2247-56.
- Giot L., Bader J. S., Brouwer C., Chaudhuri A., Kuang B., Li Y., Hao Y. L., Ooi C. E., Godwin B., Vitols E., Vijayadamar G., Pochart P., Machineni H., Welsh M., Kong Y., Zerhusen B., Malcolm R., Varrone Z., Collis A., Minto M., Burgess S., McDaniel L., Stimpson, F. Spriggs, J. Williams, K. Neurath, N. Ioime, M. Agee, E. Voss, K. Furtak, R. Renzulli E., Aanensen N., Carrola S., Bickelhaupt E., Lazovatsky Y., DaSilva A., Zhong, C. A. Stanyon, R. L. Finley, Jr., K. P. White, M. Braverman, T. Jarvie, S. Gold, M. Leach J., Knight J., Shimkets R. A., McKenna M. P., Chant J., and Rothberg J. M., "A protein interaction map of *Drosophila melanogaster*". *Science* 302.5651 (2003) 1727-36.

- Helin K., Wu C., Fattaey A. R., Lees J. A., Dynlacht B. D., Ngwu C., and Harlow E., "Heterodimerization of the transcription factors E2F-1 and DP-1 leads to cooperative trans-activation". *Genes Dev.* 7 (1993): 1850-61.
- Ito T., Tashiro K., Muta S., Ozawa R., Chiba T., Nishizawa M., Yamamoto K., Kuhara S., Sakaki Y., "Toward a protein-protein interaction map of the budding yeast: A comprehensive system to examine two-hybrid interactions in all possible combinations between the yeast proteins". *PNAS* 97.3 (2000): 1143-7.
- Kamath R. S., Fraser A. G., Dong Y., Poulin G., Durbin R., Gotta M., Kanapin A. Le Bot N., Moreno S., Sohrmann M., Welchman D. P., Zipperlen P., and Ahringer J., "Systematic functional analysis of the *Caenorhabditis elegans* genome using RNAi". *Nature* 421 (2002): 231-7.
- Kugel J. F., and Goodrich J. A., "Promoter escape limits the rate of RNA polymerase II transcription and is enhanced by TFIIE, TFIIH, and ATP on negatively supercoiled DNA". *PNAS* 95.16 (1998): 9232-37.
- Lackner M. R., Kornfeld K., Miller L. M., Horvitz R. H., and Kim S. K., "A MAP Kinase homolog, *mpk-1*, is involved in ras-mediated induction of vulval cell fates in *Caenorhabditis elegans*". *Genes Dev.* 8 (1994): 160-73.
- Laybourn P. J., and Kadonaga J. T., "Role of nucleosomal cores and histone H1 in regulation of transcription by RNA polymerase II". *Science* 254 (1991): 238-45.
- Lee, T. I., and Young R. A., "Transcription of eukaryotic protein-coding genes." *Annual Review of Genetics* 34 (2000): 77-137.
- Levine M., and Tjian R., "Transcription regulation and animal diversity". *Nature* 424 (2003): 147-52.
- Li S., Armstrong C. M., Bertin N., Ge H., Milstein S., Boxem M., Vidalain P., Han J. J., Chesneau A., Hao T., Goldberg D. S., Li N., Martinez M., Rual J., Lamesch P., Xu L., Tewari M., Wong S. L., Zhang L. V., Berriz G. F., Jacotot L., Vaglio P., Reboul J., Hirozane-Kishikawa T., Li Q., Gabel H. W., Elewa A., Baumgartner B., Rose D. J., Yu H., Bosak S., Sequerra R., Fraser A., Mango S. E., Saxton W. M., Strome S., van den Heuvel S., Piano F., Vandenhaute J., Sardet C., Gerstein M., Doucette-Stamm L., Gunsalus K. C., Harper J. W., Cusick M. E., Roth F. P., Hill D. E., and Vidal M., "A map of the interactome network of the metazoan *C. elegans*". *Science* 303.5657 (2004) 540-3.
- Mann M., Hojrup P., and Roepstorff P., "Use of mass spectrometric molecular weight information to identify proteins in sequence databases". *Biological Mass Spectrometry* 22.6 (1993): 338-45.
- Mitchell P. J., and Tjian R., "Transcriptional regulation in mammalian cells by sequence-specific DNA binding proteins". *Science* 246.4916 (1989): 371-9.
- Mizzen C. A., Yang X., Kokubo T., Brownell J. E., Bannister A. J., Owen-Hughes T., Workman J., Wang L., Berger S. L., Kouzarides T., Nakatani Y., and Allis C. D., "The TAFII250 Subunit of TFIID Has Histone Acetyltransferase Activity". *Cell* 87.7 (1996): 1261-70.
- Nakajima N., Horikoshi M., and Roeder R. G., "Factors involved in specific transcription by mammalian RNA polymerase II: purification, genetic specificity, and TATA box-promoter interaction of TFIID". *Molecular and Cellular Biology* 8.10 (1988): 4028-40.

- Noble D., "*The Music of Life Biology beyond the Genome*". New York: Oxford UP, USA, (2006): 21.
- Okuda M., Tanaka A., Araill Y., Satoh M., Okamura H., Nagadoi A., Hanaoka F., Ohkuma Y., and Nishimura Y., "A Novel Zinc Finger Structure in the Large Subunit of Human General Transcription Factor TFIIE". *J.Biol.Chem* 279.49 (2004): 51395-403.
- Ozer J., Moore P. A., Bolden A. H., Lee A., Rosen C. A., and Lieberman P. M., "Molecular cloning of the small (y) subunit of human TFIIA reveals functions critical for activated transcription". *Genes Dev.* 8 (1994): 2324-35.
- Pugh F. B., and Tjian R., "Mechanism of transcriptional activation by sp1 evidence for coactivators". *Cell* 61 (1990): 1197-207.
- Rachez C., and Freeman L. P., "Mediator complexes and transcription". *Current Opinion in Cell Biology* 13 (2001): 274-80.
- Raff J. W., Kellum R., and Alberts B., "The Drosophila GAGA transcription factor is associated with specific regions of heterochromatin throughout the cell cycle". *EMBO* 13.24 (1994): 5977-83.
- Reboul J., Vaglio P., Rual J., Lamesch P., Martinez M., Armstrong C. M., Li S., Jacotot L., Bertin N., Janky R., Troy M., Hudson J. R., Hartley J. L., Brasch M. A., Vandenhaute J., Boulton S., Endress G. A., Jenna S., Chevet E., Papatotiroopoulos V., Tolias P. P., Ptacek J., Snyder M., Huang R., Chance M. R., Lee H., Doucette-Stamm L., Hill D. E., and Vidal M., "C. elegans ORFeome version 1.1: experimental verification of the genome annotation and resource for proteome-scale protein expression". *Nature genetics* 34 (2003): 35-41
- Reece-Hoyes J., Deplancke B., Shingles J., Grove C. A., Hope I. A., and Walhout A. J. M., "A compendium of Caenorhabditis elegans regulatory transcription factors: a resource for mapping transcription regulatory networks". *Genome Biology* 6 (2005): R110.
- Robert F., Douzlech M., Forget D., Egly J., Greenblatt J., Burton Z. F., and Coulombe B., "Wrapping of Promoter DNA around the RNA Polymerase II Initiation Complex Induced by TFIIF". *Molecular Cell* 2.3 (1998): 341-51.
- Roeder R. G., "The role of general initiation factors in transcription by RNA polymerase II". *TIBS* 21 (1996): 327-35.
- Roeder R. G., "Transcriptional regulation and the role of diverse coactivators in animal cells". *FEBS* 579 (2004): 909-15.
- Rosenberg M., and Court D., "Regulatory sequences involved in the promotion and termination of RNA transcription". *Annual Review of Genetics* 13 (1979): 319-53.
- Rowland T., Baumann P., and Jackson S. P., "The TATA-binding protein: a general transcription factor in eukaryotes and archaebacteria". *Science* 264.5163 (1994): 1926-329.
- Rual J., Ceron J., Koreth J., Hao T., Nicot A., Hirozane-Kishikawa T., Vandenhaute J., Orkin S. H., Hill D. E., van den Heuvel S., and Vidal M., "Toward Improving Caenorhabditis elegans phenome mapping with an ORFeome based RNAi library". *Genome Res.* 14 (2004): 2162-8.
- Rual J., Venkatesan K., Hao T., Hirozane-Kishikawa T., Dricot A., Li N., Berriz G. F., Gibbons F. D., Dreze M., Ayivi-Guedehoussou N., Klitgord N., Simon C.,

- Boxem M., Milstein S., Rosenberg J., Goldberg D. S., Zhang L. V., Wong S. L., Franklin G., Li S., Albala J. S., Lim J., Fraughton C., Liamosas E., Cevik S., Bex C., Lamesch P., Sikorski R. S., Vandenhoute J., Zoghbi H. Y., Smolyar A., Bosak S., Sequerra R., Doucette-Stamm L., Cusick M. E., Hill D. E., Roth F. P., and Vidal M., "Towards a proteome-scale map of the protein-protein interaction network". *Nature* 437 (2005): 1173-8.
- Shin H., Hirst M., Bainbridge M. N., Magrini V., Mardis E., Moerman D. G., Marra M. A., Baillie D. L., and Jones S. J. M., "Transcriptome analysis for *Caenorhabditis elegans* based on novel expressed sequence tags". *BMC Biology* 6.30 (2008).
 - Stumpf, M. P. H., Thorne T., Silva E. D., Stewart R., An H. J., Lappe M., and Wiuf C., "Estimating the size of the human interactome". *PNAS* 105.19 (2008): 6959-64.
 - Sudarsanam P., Cao Y., Wu L., Laurent B. C., and Winston F., "The nucleosome remodeling complex, Snf/Swi, is required for the maintenance of transcription in vivo and is partially redundant with the histone acetyltransferase, Gcn5". *Embo* 18.11 (1999): 3101-06.
 - Tan S., Conaway R. C., and Conaway J. W., "Dissection of transcription factor TFIIF functional domains required for initiation and elongation". *Biochemistry* 92 (1995): 6042-46.
 - Tini M., Benecke A., Um S., Torchia J., Evans R. M., and Chambon P., "Association of CBP/p300 Acetylase and Thymine DNA Glycosylase Links DNA Repair and Transcription". *Molecular Cell* 9.2 (2002): 265-77.
 - University of Manitoba, "BIRCH, Biology Research Computer Hieracy", <http://home.cc.umanitoba.ca/~psgendb/>, 01/11/2010.
 - Verrijzer P. C., Chen J., Yokomori K., and Tjian R., "Binding of TAFs to core elements directs promoter selectivity by RNA polymerase II". *Cell* 81.7 (1995): 1115-25.
 - Zeng L., and Zhou M., "Bromodomain: an acetyl-lysine binding domain". *FEBS* 513 (2002): 124-8.
 - Zhang H., Smolen G. A., Palmer R., Christoforou A., Van den Heuvel S., and Haber D. A., "SUMO modification is required for in vivo Hox gene regulation by the *caenorhabditis elegans* polycomb group protein sop-2". *Nature Genetics* 36.5 (2004) 507-11.
 - Zheng M., Aslund F., and Storz G., "Activation of the OxyR transcription factor by reversible disulfide bond formation". *Science* 279.5357 (1998): 1718-722.

Appendix A Predicted TCFs and Phenome Data

Predicted TCFs with any form of lethality phenotype due to RNAi are shown in red.

			embryonic lethal	larval lethal	lethal
F57C7.1	Female Sterile Homeotic Protein	bromo-domain			
Y119C1B.8	tag-332	bromo-domain	√		
F13C5.2	Bromodomain Containing Protein	bromo-domain	√	√	
H20J04.2	H20J04.2	bromo-domain			
R10E11.1	cbp-1	bromo-domain, Histone Acetyltransferase	√		
F26H11.2	nurf-1	bromo-domain, PHD, ISWI/NURF Complex			
Y47G6A.6	pcaf-1	bromo-domain, Histone Acetyltransferase			√
C26C6.1	pbrm-1	bromo-domain	√	√	
F01G4.1	psa-4	bromo-domain, SWI/SNF Complex	√		
ZK783.4	flt-1	bromo-domain			
C01H6.7	tag-298	bromo-domain	√	√	
W04A8.7	taf-1	bromo-domain, TAF	√		√
F11A10.1	lex-1	bromo-domain	√		
K08H2.6	hpl-1	chromo-domain			
ZK1236.2	cec-1	chromo-domain			
K01G5.2	hpl-2	chromo-domain			
F32E10.2	Chromo-domain Containing Protein	chromo-domain	√		
T13G8.1	chd-3	PHD	√		
F26F12.7	let-418	PHD, NuRD/CHD Complex	√		
C44B9.4	athp-1	PHD			
T12D8.1	set-16	PHD, Histone Methyltransferase	√	√	
ZK593.4	rbr-2	PHD, Histone Demethylase			
F17A2.3	PHD-finger Protein	PHD			
Y59A8A.2	Y59A8A.2	PHD	√		√
K09A11.5	PHD-finger Protein	PHD			
C28H8.9	C28H8.9a	PHD			
F33E11.6	F33E11.6b	PHD			

H05L14.2	Zinc finger C3HC4 type Protein	PHD			
F42A9.2	lin-49	PHD			
C11G6.3	PHD-finger Protein	PHD			
F54F2.2	zfp-1	PHD	√		√
Y51H1A.4	ing-3	PHD			
Y37E11B.4	taf-2	TAF	√		
C11G6.1	taf-3	TAF			
R119.6	taf-4	TAF	√	√	
F30F8.8	taf-5	TAF	√		√
W09B6.2	taf-6.1	TAF			
Y37E11AL.8	taf-6.2	TAF	√		
F54F7.1	taf-7.1	TAF			
Y111B2A.16	taf-7.2	TAF			√
ZK1320.12	taf-8	TAF			
T12D8.7	taf-9	TAF	√		
K03B4.3	taf-10	TAF	√		
F48D6.1	taf-11.1	TAF			
K10D3.3	taf-11.2	TAF			
F43D9.5	taf-11.3	TAF	√		
Y56A3A.4	taf-12	TAF			
C14A4.10	taf-13	TAF			
T23C6.1	mdt-1.2	Mediator			
ZK546.13	mdt-4	Mediator	√		
Y57E12AL.5	mdt-6	Mediator	√		
Y62F5A.1	mdt-8	Mediator	√		
T09A5.6	mdt-10	Mediator	√	√	√
R144.9	mdt-11	Mediator	√	√	√
R12B2.5	mdt-15	Mediator	√		
Y113G7B.18	mdt-17	Mediator	√	√	√
C55B7.9	mdt-18	Mediator	√		
Y71H2B.6	mdt-19	Mediator	√		√
Y104H12D.1	mdt-20	Mediator			
C24H11.9	mdt-21	Mediator			
ZK970.3	mdt-22	Mediator	√		
T18H9.6	mdt-27	Mediator	√		
W01A8.1	mdt-28	Mediator			
K08E3.8	mdt-29	Mediator			√
F32H2.2	mdt-31	Mediator	√		
Y71F9B.10	sop-3, mdt-1.1	Mediator		√	
Y54E5B.3	let-49, mdt-7	Mediator	√		
F47A4.2	dpy-22, mdt-12	Mediator	√	√	

K08F8.6	let-19, mdt-13	Mediator	√		
C38C10.5	rgr-1, mdt-14	Mediator	√	√	
F39B2.4	sur-2, mdt-23	Mediator	√		
VC5.4	mys-1	Histone Acetyltransferase, TIP60/NuA4 Complex	√	√	
K03D10.3	mys-2	Histone Acetyltransferase	√		
R07B5.8	mys-3	Histone Acetyltransferase			
C34B7.4	mys-4	Histone Acetyltransferase			
C53A5.3	hda-1	Histone Deacetylase	√		
C08B11.2	hda-2	Histone Deacetylase	√	√	
Y51H1A.5	hda-3	Histone Deacetylase	√	√	√
C10E2.3	hda-4	Histone Deacetylase			
R06C1.1	hda-5	Histone Deacetylase			
F41H10.6	hdac-6	Histone Deacetylase			
C35A5.9	hdac-11	Histone Deacetylase			
T26A5.7	set-1	Histone Methyltransferase	√	√	√
C26E6.9	set-2	Histone Methyltransferase			√
C07A9.7	set-3	Histone Methyltransferase			
C32D5.5	set-4	Histone Methyltransferase	√		√
C47E8.8	set-5	Histone Methyltransferase		√	
C49F5.2	set-6	Histone Methyltransferase			
F02D10.7	set-8	Histone Methyltransferase			
F15E6.1	set-9	Histone Methyltransferase	√		
F33H2.7	set-10	Histone Methyltransferase			
F34D6.4	set-11	Histone Methyltransferase			
K09F5.5	set-12	Histone Methyltransferase			
K12H6.11	set-13	Histone Methyltransferase			
R06F6.4	set-14	Histone Methyltransferase	√	√	

R11E3.4	set-15	Histone Methyltransferase			
T21B10.5	set-17	Histone Methyltransferase			
T22A3.4	set-18	Histone Methyltransferase			
W01C8.3	set-19	Histone Methyltransferase			
W01C8.4	set-20	Histone Methyltransferase			
Y24D9A.2	set-21	Histone Methyltransferase	√		
Y32F6A.1	set-22	Histone Methyltransferase			
Y41D4B.12	set-23	Histone Methyltransferase			
Y43F11A.5	set-24	Histone Methyltransferase			
Y43F4B.3	set-25	Histone Methyltransferase	√		
Y51H4A.12	set-26	Histone Methyltransferase			
Y71H2AM.8	set-27	Histone Methyltransferase			
Y73B3B.2	set-28	Histone Methyltransferase			
Y92H12BR.6	set-29	Histone Methyltransferase			
ZC8.3	set-30	Histone Methyltransferase			
C15H11.5	set-31	Histone Methyltransferase			
C41G7.4	set-32	Histone Methyltransferase			
Y108F1.3	set-33	Histone Methyltransferase	√		
K07C11.2	air-1	Histone Kinase	√		√
B0207.4	air-2	Histone Kinase	√		
F29B9.6	ubc-9	Histone Summoylase	√		√
T08D10.2	lsd-1	Histone Demethylase	√	√	√
Y40B1B.6	spr-5	Histone Demethylase			
T26A5.5	T26A5.5	Histone Demethylase			√
F29B9.2	F29B9.2	Histone Demethylase	√		
F43G6.6	F43G6.6	Histone Demethylase			
F29B9.4	psr-1	Histone Demethylase			

T07C4.11	T07C4.11	Histone Demethylase	√		√
Y48B6A.11	jmjd-2	Histone Demethylase			
D2021.1	utx-1	Histone Demethylase	√		√
F18E9.5	tag-279	Histone Demethylase			
C29F7.6	C29F7.6	Histone Demethylase			
Y2H9A.1	mes-4	SET-domain			√
R06A4.7	mes-2	SET-domain			
C43E11.3	met-1	SET-domain			
R05D3.11	met-2	SET-domain			√
T12F5.4	lin-59	SET-domain		√	√
C18E3.2	C18E3.2	SWI/SNF Complex	√		√
F26D10.3	hsp-1	SWI/SNF Complex	√		√
C01G8.9	let-526	SWI/SNF Complex	√	√	
Y113G7B.23	psa-1	SWI/SNF Complex		√	√
R07E5.3	snfc-5	SWI/SNF Complex		√	
Y111B2A.22	ssl-1	SWI/SNF Complex		√	√
B0041.7	xnp-1	SWI/SNF Complex			
Y71H2AM.17	Y71H2AM.17	SWI/SNF Complex	√	√	
ZK1128.5	tag-246	SWI/SNF Complex		√	√
ZK616.4	ZK616.4	SWI/SNF Complex	√		
K07A1.12	lin-53	NuRD/CHD Complex	√	√	√
M04G2.1	mep-1	NuRD/CHD Complex		√	√
F37A4.8	isw-1	ISWI/NURF Complex	√	√	√
C47E12.4	pyp-1	ISWI/NURF Complex	√	√	
K07A1.11	rba-1	ISWI/NURF Complex	√		√
C08B11.6	C08B11.6	SWR1/SRCAP Complex	√		
C17E4.6	C17E4.6	SWR1/SRCAP Complex	√		
CD4.7	CD4.7	SWR1/SRCAP Complex			
M04B2.3	gfl-1	SWR1/SRCAP	√		

		Complex			
Y37D8A.9	mrg-1	SWR1/SRCAP Complex			
R08C7.3	htz-1	SWR1/SRCAP Complex	√	√	√
Y105E8A.17	ekl-4	SWR1/SRCAP Complex	√		√
C14B1.4	swd-3.1	COMPASS Complex			
ZK863.6	dpy-30	COMPASS Complex	√	√	
C46A5.9	hcf-1	COMPASS Complex			
Y53G8AR.2	Y53G8AR.2	NuA3 Complex			
Y111B2A.11	epc-1	TIP60/NuA4 Complex	√		
C47D12.1	trr-1	TIP60/NuA4 Complex	√	√	
ZK1127.3	ZK1127.3	TIP60/NuA4 Complex			

Appendix B Blast Result Alignments

Appendix B.3 Lsd-1 Spr-5 Alignment

>Y40B1B.6 CE20240 WBGene00005010 locus:spr-
5#status:Confirmed#UniProt:Q9XWP6#protein_id:CAA21604.1
Length = 770

Score = 520 bits (1339), Expect = e-147, Method: Composition-based stats.
Identities = 281/650 (43%), Positives = 404/650 (62%), Gaps = 12/650 (1%)

Query: 82 DRPTEIEAAFFPEVQMSRSFSDVFLMIRNTTLSIWLASATTECTAEDVIKHLTPPYNTEI 141
DRPT+ E AFFPE+ ++ +VFL++RN+TL+ W + ECTA DV ++ PP+N+++
Sbjct: 40 DRPTDHELAFPELWEHKTAVEVFLLLRNSTLATWQYNPLKECTALDVRNNVFPFNSDL 99

Query: 142 HLVQNIVLFLSRFGMINIGFFFPKTELVNM--EKKFXXXXXXXXXXXXXTQLLTFGFD 199
L+QNIV +LSR G+IN G + T++ + +++ TQL +FGFD
Sbjct: 100 DLIQNIVHYLSRHGLINFGRYVRSTKISRFLVRDRRSVIVIGAGAAGISAATQLESFGFD 159

Query: 200 VAVVEASGLTGGRVRSLSKKGELIETGCDSLRNLDSEVITLLHQVPLNENIMSNTIV 259
V V+EA GGR+ S SK GE++ETG D+LR +++S + TLLHQV E+ + + T V
Sbjct: 160 VIVLEARNICGGRIHSFKSKSGEIMETGGDTLRKIEDSPMATLLHQVNFEEHGVDFDTSV 219

Query: 260 FSKGKYVPVARCHVINGLYANLKAAGLAHASHGPEQGENGLYISRQQAYENYFNMIERST 319
F +G+ + + H+ Y + L + +H E R + G +ISRQQAYEN +M ER T
Sbjct: 220 FVEGRPLNEEKIHLFLDHYKSAHGALNYQAHQCEHRDDQGSFISRQQAYENLLSMCERG 279

Query: 320 LLSYYNFAKEKVNLNAERKHYEVLKTNRLTALLAEQKLKNTTP-----SDELLKSLQI 374
L+ YYNF K + R+H + +K R+TAL+AE +LK D +L +SL+
Sbjct: 280 LIKYNFCKSLETVARAREHHFNQMKQLRMTALMAENQLKMEEEGNLEQDPVLRSLKR 339

Query: 375 DIEKAIKRFDEACERFEICEERADLEKNPRCKQSMHP-NDFIHYNFLLGFEERLFGAQL 433
DI ++ +F+E + FE + L ++P+ KQ MHP ++F +NF+LGFE L GAQL
Sbjct: 340 DIATSLEKFEEVADAFETADNHWQRLNEHPQAKQMHPGSEFATFNFMGLGFEEYLVAQL 399

Query: 434 EKVQFSCNVNELKLSQVARVQEGLAQVLIINVANERKVKIHHNQRVIEIDTGSSDAVILK 493
EKVQFSC+ + K AR+ EG+A++L ++ +RK+ I RV++ID + V+LK
Sbjct: 400 EKVQFSCDSMQNKENGVAARLTEGIAELLTQLSEKRKLDIRLKHRLDIDYSGFEHVLLK 459

Query: 494 LRKPDGSVGILNADYVVSTLPIGVLLKTIIGDERAPVFRPPLPKSKFAAIRSLGNLINK 553
+++ +G + + A +VVSTLPIGVLLKTTI DERAP F P LP K AIR++G G +NK
Sbjct: 460 VQRENGDIEEMKAFFVSTLPIGVLLKTTIADERAPFTFSLPDKKVEAIRNIGCGSVNK 519

Query: 554 IVFVFETRFPES--INQFAIVPDKISERAAMFTWSSLPESTRLTTHYVGENRFHDTPT 611
+ F+ FW + NQF V I R +M WSS+P S+ L T+ VGE + P
Sbjct: 520 CILEFDRVFWTANGGRNQFVTVSPNIKTRGSMNIWSSVPGSKVLCTYIVGEEAMLELPDD 579

Query: 612 ELITKALEMLKTVF-KDCP-SPIDAYVTNWHTDELAFGTGFMSLRTEPQHFDALKEPLK 669
+I A+ L+ F +CP +PI A++T WH DELAFG+G FMSLRTE FD + EPLK
Sbjct: 580 VIIQNAMINLQKAFGNCCPRAPISAHITRWHDDELAFGSGAFMSLRTEPQFDVMEPLK 639

Query: 670 TRDGKPRVFFAGEHTSALEHGTLDGAFNSGLRAAADLANTCIEIPFINRS 719
T DG RV+PAGEHT + T+ GA+ SG RAAAD++N I I F++ S
Sbjct: 640 TSDGMSRVYFAGEHTCSSYTSTIQGAWMSGARAAADISNDHIGIGFVDIS 689

Appendix B.2 Bromo-domain BLAST Alignment

>F57C7.1a CE31548 WBGene00010199 female sterile homeotic protein
(Bromodomain
protein)#status:Partially_confirmed#UniProt:Q20947#prote
in_id:CAA93473.3
Length = 1209

Score = 124 bits (312), Expect = 8e-30, Method: Composition-based stats.
Identities = 55/110 (50%), Positives = 77/110 (70%)

Query: 1 PKRQTNQLQYLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLE 60
P R TN L ++L V+K KH+ +WPFQ PVDA+KL +P+Y+ I+ TPMD+ TI+KRL
Sbjct: 280 PTRHTNCLDFVLFTVVKDALKHKHSWPFQLPVDAIKLEIPEYHNIVNTPMDLRTIEKRRL 339

Query: 61 NNYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKINELP 110
N YYW A++ I+D N +F NCY +N P D+ MA+ LEK L ++ +LP
Sbjct: 340 NLYWCAEDAIAKDINQVFINCYSFNPPEYDVYKMAKTLEKQVLSQLTQLP 389

Score = 62.8 bits (151), Expect = 3e-11, Method: Composition-based stats.
Identities = 34/87 (39%), Positives = 44/87 (50%)

Query: 24 FAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLENNYYWNAQECIQDFNTMFTNCYI 83
FA F PVD +KL + DY ++I PMD+ TIKK+L+ Y +E + D N M NC
Sbjct: 575 FAQVFYLPVDPIKLIYDYLEVITNPMDLQTIKKKLDKQYAEPEEFVHDINLMVDNCK 634

Query: 84 YNKPGGDIVLMAEAEKLFQKINELP 110
YN G A L F Q+ P
Sbjct: 635 YNPKGSPAHSNALELRSFQEQRWKLFP 661

>F57C7.1b CE18761 WBGene00010199 female sterile homeotic protein
(Bromodomain
protein)#status:Partially_confirmed#UniProt:Q20948#prote
in_id:CAA93475.1
Length = 1087

Score = 119 bits (299), Expect = 2e-28, Method: Composition-based stats.
Identities = 52/111 (46%), Positives = 76/111 (68%)

Query: 1 PKRQTNQLQYLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLE 60
P R TN L ++L V+K KH+ +WPFQ PVDA+KL +P+Y+ I+ TPMD+ TI+KRL
Sbjct: 280 PTRHTNCLDFVLFTVVKDALKHKHSWPFQLPVDAIKLEIPEYHNIVNTPMDLRTIEKRRL 339

Query: 61 NNYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKINELPT 111
N YYW A++ I+D NT+F NC +N DDI +M E +E + + + +P+
Sbjct: 340 NLYWCAEDAIAKDLNLFNCKKFNDRNDDIYIMCENIEGVVQRGLEWMP 390

Score = 63.5 bits (153), Expect = 2e-11, Method: Composition-based stats.
Identities = 34/87 (39%), Positives = 44/87 (50%)

Query: 24 FAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLENNYYWNAQECIQDFNTMFTNCYI 83
FA F PVD +KL + DY ++I PMD+ TIKK+L+ Y +E + D N M NC
Sbjct: 575 FAQVFYLPVDPIKLIYDYLEVITNPMDLQTIKKKLDKQYAEPEEFVHDINLMVDNCK 634

Query: 84 YNKPGGDIVLMAEAEKLFQKINELP 110
YN G A L F Q+ P
Sbjct: 635 YNPKGSPAHSNALELRSFQEQRWKLFP 661

>Y119C1B.8a CE44037 WBGene00022473 locus:tag-
332#status:Partially_confirmed#UniProt:Q95Y80#protein_id
:AAK39326.3
Length = 853

Score = 114 bits (286), Expect = 7e-27, Method: Composition-based stats.
Identities = 49/110 (44%), Positives = 73/110 (66%)

Query: 1 PKRQTNQLQYLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLE 60
P R TN+L Y++ VLK KH+ WPFQ+PVDAV L +P Y++ + PMD+ TI+ RL+
Sbjct: 37 PTRHTNKLDYIMTTVLKEAGKHKHVWPFQKPVDAVALCIPLYHERVARPMDLKTIENTRLK 96

Query: 61 NNYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEALEKLFQKINELP 110
+ YY AQECI D T+F NCY +N DD+ +MA+ + ++ + + + P
Sbjct: 97 STYYTCAQECIDDIEYTFQNCYTFNGKEDDVTIMAQNVHEVIKKSLEQAP 146

Score = 76.3 bits (186), Expect = 3e-15, Method: Composition-based stats.
Identities = 34/88 (38%), Positives = 53/88 (60%)

Query: 22 HQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLENNYWNAQECIQDFNTMFTNC 81
+FAWPF +PVDA +L L DY+KIIK PMD+ ++K ++E+ Y + D M NC
Sbjct: 279 QEFAWPFNEPVDAEQLGLHDYHKIIEKPMDLKSMKAKMESGAYKEPSDFEHDVRLMLRNC 338

Query: 82 YIYNKPGDDIVLMAEALEKLFQKINEL 109
++YN GD + +++F ++ EL
Sbjct: 339 FLYNPVGDVHVSFGLRFQEVFDRRWAE 366

>Y119C1B.8b CE33207 WBGene00022473 locus:tag-
332#status:Partially_confirmed#UniProt:Q86S79#protein_id
:AAO21405.1
Length = 765

Score = 114 bits (285), Expect = 1e-26, Method: Composition-based stats.
Identities = 49/110 (44%), Positives = 73/110 (66%)

Query: 1 PKRQTNQLQYLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLE 60
P R TN+L Y++ VLK KH+ WPFQ+PVDAV L +P Y++ + PMD+ TI+ RL+
Sbjct: 37 PTRHTNKLDYIMTTVLKEAGKHKHVWPFQKPVDAVALCIPLYHERVARPMDLKTIENTRLK 96

Query: 61 NNYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEALEKLFQKINELP 110
+ YY AQECI D T+F NCY +N DD+ +MA+ + ++ + + + P
Sbjct: 97 STYYTCAQECIDDIEYTFQNCYTFNGKEDDVTIMAQNVHEVIKKSLEQAP 146

Score = 76.3 bits (186), Expect = 3e-15, Method: Composition-based stats.
Identities = 34/88 (38%), Positives = 53/88 (60%)

Query: 22 HQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLENNYWNAQECIQDFNTMFTNC 81
+FAWPF +PVDA +L L DY+KIIK PMD+ ++K ++E+ Y + D M NC
Sbjct: 279 QEFAWPFNEPVDAEQLGLHDYHKIIEKPMDLKSMKAKMESGAYKEPSDFEHDVRLMLRNC 338

Query: 82 YIYNKPGDDIVLMAEALEKLFQKINEL 109
++YN GD + +++F ++ EL
Sbjct: 339 FLYNPVGDVHVSFGLRFQEVFDRRWAE 366

>F13C5.2 CE19384 WBGene00017423 bromodomain-containing
protein#status:Confirmed#UniProt:O76561#protein_id:AAC64
610.1
Length = 374

Score = 78.2 bits (191), Expect = 8e-16, Method: Composition-based stats.
Identities = 37/82 (45%), Positives = 47/82 (57%)

Query: 24 FAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRLENNYWNAQECIQDFNTMFTNCYI 83
F +PF++PVD V L L DY+++IK PMDM TI+K+L Y A E +DF M NC
Sbjct: 137 FTFFPRKPVDDVLLGLTDYHEVIKPKMDMSTIRKKLIGEEYDTAVEFKEDFKLMINNCLT 196

Query: 84 YNKPDDIVLMAEALEKLFQK 105

YN GD + A K F K
Sbjct: 197 YNNEGDPVADFALQFRKKFAAK 218

>H20J04.2 CE27187 WBGene00019217 status:Partially_confirmed UniProt:Q9
N5L9#protein_id:AAF39888.2
Length = 1427

Score = 70.5 bits (171), Expect = 2e-13, Method: Composition-based stats.
Identities = 31/99 (31%), Positives = 57/99 (57%), Gaps = 2/99 (2%)

Query: 11 LLRVVLTWLVKHWQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNYYWNAQEC 70
L+ +LK + +WPF QPVD+ ++ PDYY +IK PM++ T+ +++ Y E
Sbjct: 1328 LIETLLKEAMRQEC5WPFLQPVD5KEV--PDYYDVIKRPMLNLTMMNKIKQRIYNKPIEV 1385

Query: 71 IQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKINEL 109
DF + +NC YN+P ++I ++ L +++E+
Sbjct: 1386 RNFQQLILSNCEYNEPENEIYKLSRELHDFMADRLDEI 1424

>R10E11.1c CE42151 WBGene00000366 locus:cbp-
1#status:Partially_confirmed#UniProt:B0M0M3#protein_id:C
AP72377.1
Length = 2016

Score = 68.9 bits (167), Expect = 4e-13, Method: Composition-based stats.
Identities = 38/104 (36%), Positives = 61/104 (58%), Gaps = 1/104 (0%)

Query: 4 QTNQLQYLLRVVLTWLVKHWQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNY 63
Q + +++LL V K L K + A PF+ PVDA LN+PDY++IIK PMD+ T+ K+L
Sbjct: 855 QEDLIKFLLPVWEK-LDKSEDAAPFRVPVDAKLLNIPDYHEIIKRPMDLETVHKKLYAGQ 913

Query: 64 YWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKIN 107
Y NA + D M N ++YN+ + L ++F+ +++
Sbjct: 914 YQNAQGFCDIWLMLDNAWLYNRKNSKVYKYGLKLSMFVSEMD 957

>R10E11.1b CE21117 WBGene00000366 locus:cbp-
1#status:Partially_confirmed#UniProt:P34545#protein_id:C
AD18875.1
Length = 2056

Score = 68.9 bits (167), Expect = 4e-13, Method: Composition-based stats.
Identities = 38/104 (36%), Positives = 61/104 (58%), Gaps = 1/104 (0%)

Query: 4 QTNQLQYLLRVVLTWLVKHWQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNY 63
Q + +++LL V K L K + A PF+ PVDA LN+PDY++IIK PMD+ T+ K+L
Sbjct: 866 QEDLIKFLLPVWEK-LDKSEDAAPFRVPVDAKLLNIPDYHEIIKRPMDLETVHKKLYAGQ 924

Query: 64 YWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKIN 107
Y NA + D M N ++YN+ + L ++F+ +++
Sbjct: 925 YQNAQGFCDIWLMLDNAWLYNRKNSKVYKYGLKLSMFVSEMD 968

>R10E11.1a CE28069 WBGene00000366 locus:cbp-
1#bromodomain#status:Partially_confirmed#UniProt:P34545#
protein_id:CAA82353.2
Length = 2045

Score = 68.9 bits (167), Expect = 4e-13, Method: Composition-based stats.
Identities = 38/104 (36%), Positives = 61/104 (58%), Gaps = 1/104 (0%)

Query: 4 QTNQLQYLLRVVLTWLVKHWQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNY 63
Q + +++LL V K L K + A PF+ PVDA LN+PDY++IIK PMD+ T+ K+L

Sbjct: 855 QEDLIKFLLPVWEK-LDKSEDAAPFRVPVDAKLLNIPDYHEIIKRPMDLETVHKKLYAGQ 913

Query: 64 YWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAEKLFQKIN 107
Y NA + D M N ++YN+ + L ++F+ +++

Sbjct: 914 YQNAQGFCDDIWLMLDNAWLYNRKNSKVYKYGLKLEMFVSEMD 957

>F26H11.2e CE15909 WBGene00009180 locus:nurf-
1#Bromodomain#status:Confirmed#UniProt:Q6BER5#protein_id
:CAB04198.1
Length = 405

Score = 67.4 bits (163), Expect = 2e-12, Method: Composition-based stats.
Identities = 34/86 (39%), Positives = 50/86 (58%), Gaps = 4/86 (4%)

Query: 22 HQFAWPFQQPVDVAVKLN-LPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTN 80
H+ + PF+ PVD LN PDY K IK PMD+ TI K++E Y + + D N MF N

Sbjct: 260 HRMSTPFRNPVD---LNEFPDYEKFIKPPMDLSTITKKVERTEYLYLSQFVNDVNQMFEN 316

Query: 81 CYIYNKPGDDIVLMAEAEKLFQKI 106
YN G+ + AE ++++F +K+

Sbjct: 317 AKTYNPKGNAVFKCAETMQEVFDKKL 342

>F26H11.2f CE15910 WBGene00009180 locus:nurf-
1#Bromodomain#status:Confirmed#UniProt:Q6BER5#protein_id
:CAB04195.1
Length = 510

Score = 67.0 bits (162), Expect = 2e-12, Method: Composition-based stats.
Identities = 34/86 (39%), Positives = 50/86 (58%), Gaps = 4/86 (4%)

Query: 22 HQFAWPFQQPVDVAVKLN-LPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTN 80
H+ + PF+ PVD LN PDY K IK PMD+ TI K++E Y + + D N MF N

Sbjct: 365 HRMSTPFRNPVD---LNEFPDYEKFIKPPMDLSTITKKVERTEYLYLSQFVNDVNQMFEN 421

Query: 81 CYIYNKPGDDIVLMAEAEKLFQKI 106
YN G+ + AE ++++F +K+

Sbjct: 422 AKTYNPKGNAVFKCAETMQEVFDKKL 447

>F26H11.2d CE42388 WBGene00009180 locus:nurf-
1#status:Confirmed#UniProt:Q6BER5#protein_id:CAB54234.4
Length = 808

Score = 66.6 bits (161), Expect = 2e-12, Method: Composition-based stats.
Identities = 34/86 (39%), Positives = 50/86 (58%), Gaps = 4/86 (4%)

Query: 22 HQFAWPFQQPVDVAVKLN-LPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTN 80
H+ + PF+ PVD LN PDY K IK PMD+ TI K++E Y + + D N MF N

Sbjct: 663 HRMSTPFRNPVD---LNEFPDYEKFIKPPMDLSTITKKVERTEYLYLSQFVNDVNQMFEN 719

Query: 81 CYIYNKPGDDIVLMAEAEKLFQKI 106
YN G+ + AE ++++F +K+

Sbjct: 720 AKTYNPKGNAVFKCAETMQEVFDKKL 745

>F26H11.2g CE37638 WBGene00009180 locus:nurf-
1#status:Confirmed#UniProt:Q6BER5#protein_id:CAH60782.1
Length = 413

Score = 66.6 bits (161), Expect = 2e-12, Method: Composition-based stats.
Identities = 34/86 (39%), Positives = 50/86 (58%), Gaps = 4/86 (4%)

Query: 22 HQFAWPFQQPVDVAVKLN-LPDYKIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTN 80
H+ + PF+ PVD LN PDY K IK PMD+ TI K++E Y + + D N MF N
Sbjct: 268 HRMSTPFRNPVD---LNEFPDYEFIKKPMDLSTITKKVERTEYLYLSQFVNDVNMFMEN 324

Query: 81 CYIYNKPGDDIVLMAEAEKLFQKI 106
YN G+ + AE ++++F +K+
Sbjct: 325 AKTYNPKGNAVFKAETMQEVFDKKL 350

>F26H11.2c CE36931 WBGene00009180 locus:nurf-
1#status:Partially_confirmed#UniProt:Q6BER5#protein_id:CA
H04722.1
Length = 2266

Score = 66.2 bits (160), Expect = 3e-12, Method: Composition-based stats.
Identities = 34/86 (39%), Positives = 50/86 (58%), Gaps = 4/86 (4%)

Query: 22 HQFAWPFQQPVDVAVKLN-LPDYKIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTN 80
H+ + PF+ PVD LN PDY K IK PMD+ TI K++E Y + + D N MF N
Sbjct: 2121 HRMSTPFRNPVD---LNEFPDYEFIKKPMDLSTITKKVERTEYLYLSQFVNDVNMFMEN 2177

Query: 81 CYIYNKPGDDIVLMAEAEKLFQKI 106
YN G+ + AE ++++F +K+
Sbjct: 2178 AKTYNPKGNAVFKAETMQEVFDKKL 2203

>Y47G6A.6 CE24372 WBGene00021636 locus:pcaf-
1#status:Partially_confirmed#UniProt:Q9N3S7#protein_id:A
AF60658.1
Length = 767

Score = 63.9 bits (154), Expect = 2e-11, Method: Composition-based stats.
Identities = 35/95 (36%), Positives = 53/95 (55%), Gaps = 10/95 (10%)

Query: 15 VLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIKTPMDMGTIKKRENNYYWNAQECIQDF 74
+LK L + AWP PVD ++ P+YY IK P+D T++++L+ Y + I D
Sbjct: 658 ILKKLTADKNAWPFASPVVDVKEV--PEYYDHIKHPIDFKTMQEKLKRKAYTHQHLFIADL 715

Query: 75 NTMFTNCYIYNKPGDDIVLMAEAEKLFQKINEL 109
N +F NCY++N AEA+ + K+NEL
Sbjct: 716 NRLFQNCYVFNG-----AEAVYYKYGYKLNEL 742

>C26C6.1a CE30254 WBGene00007042 locus:pbrm-1 HMG (high mobility
group) box, Bromodomain (5 domains), Zinc finger, C2H2
type#status:Partially_confirmed#UniProt:Q18210#protein_i
d:CAA96697.2
Length = 1883

Score = 55.8 bits (133), Expect = 4e-09, Method: Composition-based stats.
Identities = 30/76 (39%), Positives = 40/76 (52%)

Query: 36 KLNLPDYKIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMA 95
K P YY +IK PMDM IK +LEN Y + + DF M +N +N+ DI A
Sbjct: 743 KEEFPAYYDVIKPMDDMRIRKHLNRYVTLDDVVSDFMLMLSNACKFNEDSDIYKEA 802

Query: 96 EALEKLFQKINELPT 111
+L+K L+ EL T
Sbjct: 803 VSLQKALLEMKRELDLPT 818

Score = 50.4 bits (119), Expect = 2e-07, Method: Composition-based stats.
Identities = 25/65 (38%), Positives = 38/65 (58%)

Query: 40 PDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAL 99
P+YY+II+ P+DM TI+ R++ + Y I D MF+N +N+P I + A LE
Sbjct: 570 PEYYQIIQNPIDMKTIRMIRIDGHQYPQVDAMINDCRVFMFSNARDFNEPRSMIHMDAIQLE 629

Query: 100 KLFLQ 104
K L+
Sbjct: 630 KAVLR 634

Score = 46.2 bits (108), Expect = 4e-06, Method: Composition-based stats.
Identities = 24/74 (32%), Positives = 40/74 (54%), Gaps = 1/74 (1%)

Query: 36 KLNLPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMA 95
K + PDYY IK P+ + I KRL+N Y + + + D M++N + YN ++ + A
Sbjct: 374 KESYPDYYDEIKNPVSIFMINKRLKNGKY-DLKSLVADLMQMYNAFDYNLESSEVYISA 432

Query: 96 EALEKLFLQKINEL 109
E L+ L + +L
Sbjct: 433 EKLKALTISTCKQL 446

Score = 39.3 bits (90), Expect = 4e-04, Method: Composition-based stats.
Identities = 23/72 (31%), Positives = 35/72 (48%)

Query: 38 NLPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEA 97
+ P YY+ I P+D+ TI+ N Y +E D +F N ++ G DI AE
Sbjct: 226 DFPLYEYKIAKPIDLKTIAQNGVNKKYSTMKELKDDLFLFKNAQQFSGNGSDIFKDAEQ 285

Query: 98 LEKLFLQKINEL 109
L+ + +KI L
Sbjct: 286 LKTVVKEKIARL 297

Score = 31.2 bits (69), Expect = 0.12, Method: Composition-based stats.
Identities = 21/70 (30%), Positives = 32/70 (45%), Gaps = 8/70 (11%)

Query: 40 PDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAL 99
P+YY+ +K P+D+ TI+ +L+ Y + DF N Y K E+ E
Sbjct: 79 PEYEQVKEPIDVTTIQHKLKIPEYLTYDQFNDDFMMFIKNNLTYYKD-----ESEE 130

Query: 100 KLFLQKINEL 109
+ KI EL
Sbjct: 131 HKDMMKIQEL 140

>F01G4.1 CE05553 WBGene00004204 locus:psa-4 SNF2alpha
like#status:Confirmed#UniProt:Q19106#protein_id:CAA92978.
1
Length = 1474

Score = 52.4 bits (124), Expect = 5e-08, Method: Composition-based stats.
Identities = 24/71 (33%), Positives = 40/71 (56%)

Query: 39 LPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKPGDDIVLMAEAL 98
LPDYY++I PMD I K++E Y +E D N + N YN+ G +I + +E +
Sbjct: 1217 LPDYYQVISKPMDFDRINKKIETGRYTVMEELNDDMNLVNNNAQTYNEEGSEIYVSSETI 1276

Query: 99 EKLFLQKINEL 109
KL+ ++ ++
Sbjct: 1277 GKLWKEQYDKF 1287

>ZK783.4 CE34152 WBGene00001470 locus:flt-
1#status:Partially_confirmed#UniProt:Q23590#protein_id:AA
C24421.2
Length = 1376

Score = 49.7 bits (117), Expect = 3e-07, Method: Composition-based stats.
Identities = 33/104 (31%), Positives = 53/104 (50%), Gaps = 2/104 (1%)

Query: 6 NQLQYLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNYYW 65
N + L +++L L A PF +PV+ KL +P Y II PMD+ TI+++ E Y
Sbjct: 1262 NMNKELCQLMLDELVVQANALPFLEPVNP-KL-VPGYKMIISKPMDLKTIRQKNEKLIYE 1319

Query: 66 NAQECIQDFNTMFTNCYIYNKPGDDIVLMAEALEKLFLQKINEL 109
++ +D MF NC +N +I +L K F ++ +L
Sbjct: 1320 TPEDFAEDIELMFANCRQFNIDHSEIGRAGISLHKFFQKRWKQL 1363

>C01H6.7a CE05190 WBGene00007256 locus:tag-
298#Bromodomain#status:Confirmed#UniProt:Q17581#protein_
id:CAA95779.1
Length = 636

Score = 48.1 bits (113), Expect = 9e-07, Method: Composition-based stats.
Identities = 28/92 (30%), Positives = 48/92 (52%), Gaps = 6/92 (6%)

Query: 10 YLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNYYWNAQE 69
++LR +++ + FA+P + PDY IIKTPMD+ TI++ +E+ Y +
Sbjct: 158 HILRKLVEKDPEQYFAFPVTPSM-----APDYRDIKTPMDLQTIRENIEDGKYASLPA 211

Query: 70 CIQDFNTMFTNCYIYNKPGDDIVLMAEALEKL 101
+D + +N + YN+P L A+ L L
Sbjct: 212 MKEDCELIVSNAFQYNQPNTVFYLAARKLSNL 243

>C01H6.7b CE40891 WBGene00007256 locus:tag-
298#status:Confirmed#UniProt:A5JYT2#protein_id:CAN86573.
1
Length = 582

Score = 47.4 bits (111), Expect = 1e-06, Method: Composition-based stats.
Identities = 28/92 (30%), Positives = 48/92 (52%), Gaps = 6/92 (6%)

Query: 10 YLLRVVLKTLWKHQFAWPFQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNYYWNAQE 69
++LR +++ + FA+P + PDY IIKTPMD+ TI++ +E+ Y +
Sbjct: 158 HILRKLVEKDPEQYFAFPVTPSM-----APDYRDIKTPMDLQTIRENIEDGKYASLPA 211

Query: 70 CIQDFNTMFTNCYIYNKPGDDIVLMAEALEKL 101
+D + +N + YN+P L A+ L L
Sbjct: 212 MKEDCELIVSNAFQYNQPNTVFYLAARKLSNL 243

>W04A8.7 CE42634 WBGene00006382 locus:taf-1 transcription initiation
factor
TFIID#status:Partially_confirmed#UniProt:Q9XUL9#protein_i
d:CAC14425.2
Length = 1744

Score = 44.3 bits (103), Expect = 1e-05, Method: Composition-based stats.
Identities = 25/81 (30%), Positives = 44/81 (54%), Gaps = 2/81 (2%)

Query: 28 FQQPVDVAVKLNLPDYKIIKTPMDMGTIKKRENNYYWNAQECIQDFNTMFTNCYIYNKP 87
F PV++ K+ DYY IIK P+ + IKK++ Y ++ + D MF N +YN

Sbjct: 1433 FVTPVNSKKV--VDYYNIIKNPISLQEIKKKISEQSYLLRKDFLDDIKLMFDNSRMVNGD 1490

Query: 88 GDDI¹VLMAEALEKFLQKINE 108
+ + L A + + L +++ E

Sbjct: 1491 NNILTLTAQQLQLAGKRMIE 1511

Score = 36.6 bits (83), Expect = 0.002, Method: Composition-based stats.
Identities = 31/119 (26%), Positives = 55/119 (46%), Gaps = 12/119 (10%)

Query: 1 PKRQTNQL--QYLLRVVLKTLWKHQFAWPFQQP¹DAVKLNLPD¹YKIIKTPMDMGTIKK 57
P TN L YLL +++ + + F VD K+ P YY I PMD+ +++

Sbjct: 1525 PLLDNTDLIGFSYLLGEIVQMKMKNPKSALFHTRVDPKKI--PAYYLKISDPMDLISIMEQ 1582

Query: 58 RLENNYYWNAQECIQDFNTMFTNCYI¹YNKP-----GDDI¹VLMAEALEKFLQKINEL 109
+ ++ Y + E ++D ++TN ++N ++ MAE L K + + EL

Sbjct: 1583 KSKSQEYKSIDFLKDAEKIYTNVVFNGAESVYSLKAKEMFEMAEMLVKQMDTLGEL 1641

>F11A10.1c CE20665 WBGene00008682 locus:lex-
1#status:Confirmed#UniProt:P54816#protein_id:CA082045.1
Length = 1242

Score = 40.8 bits (94), Expect = 1e-04, Method: Composition-based stats.
Identities = 35/112 (31%), Positives = 52/112 (46%), Gaps = 16/112 (14%)

Query: 1 PKRQTNQLQYLLRVVLKT-----LWKHQFAWPFQQP¹DAVKLNLPD¹YKIIKTPM 50
P R+T + +Y V+ K L + + F +PVD + DYY+II+TP+

Sbjct: 853 PSRRIRQKYFEHVIEKINTPPKVFDPRLMRDRRFVEFVEPVPDPEAE--DYEIIETPI 910

Query: 51 DMGTIKKRLENNYYWNAQECIQDFNTMFTNCYI¹YN----KPGDDI¹VLMAEAL 98
M I ++L N Y +A + + D + TN YN K G I MA L

Sbjct: 911 CMQDIMEKLNCEYNHADKFVADLILIQTNALEYNPSTTKDGKLRQMANTL 962

>F11A10.1b CE41384 WBGene00008682 locus:lex-
1#status:Confirmed#UniProt:P54816#protein_id:CA082044.1
Length = 1289

Score = 40.4 bits (93), Expect = 2e-04, Method: Composition-based stats.
Identities = 31/96 (32%), Positives = 47/96 (48%), Gaps = 6/96 (6%)

Query: 7 QLQYLLRVVLKTLWKHQFAWPFQQP¹DAVKLNLPD¹YKIIKTPMDMGTIKKRLENNYYWN 66
Q++ + L L + + F +PVD + DYY+II+TP+ M I ++L N Y +

Sbjct: 916 QMRLFFKERLTRLMRDRRFVEFVEPVPDPEAE--DYEIIETPICMQDIMEKLNCEYNH 973

Query: 67 AQECIQDFNTMFTNCYI¹YN----KPGDDI¹VLMAEAL 98
A + + D + TN YN K G I MA L

Sbjct: 974 ADKFVADLILIQTNALEYNPSTTKDGKLRQMANTL 1009

>F11A10.1a CE40608 WBGene00008682 locus:lex-1 TAT-binding homolog
like#status:Confirmed#UniProt:P54816#protein_id:CAA92684.
2
Length = 1291

Score = 40.4 bits (93), Expect = 2e-04, Method: Composition-based stats.
Identities = 31/96 (32%), Positives = 47/96 (48%), Gaps = 6/96 (6%)

Query: 7 QLQYLLRVVLKTLWKHQFAWPFQQP¹DAVKLNLPD¹YKIIKTPMDMGTIKKRLENNYYWN 66
Q++ + L L + + F +PVD + DYY+II+TP+ M I ++L N Y +

Sbjct: 918 QMRLFFKERLTRLMRDRRFVEFVEPVPDPEAE--DYEIIETPICMQDIMEKLNCEYNH 975

Query: 67 AQECIQDFNTMFTNCYI¹YN----KPGDDI¹VLMAEAL 98

A + + D + TN YN K G I MA L
Sbjct: 976 ADK FVADLILIQTNALEYNPSTTKDGLIRQMANTL 1011

>F26H11.2h CE42387 WBGene00009180 locus:nurf-
1#status:Confirmed#UniProt:Q6BER5#protein_id:CAQ16138.1
Length = 554

Score = 30.4 bits (67), Expect = 0.20, Method: Composition-based stats.
Identities = 15/52 (28%), Positives = 29/52 (55%), Gaps = 1/52 (1%)

Query: 55 IKKRLNNYYWNAQECIQDFNTMFTNCYIYNKPGDDI VLMAEAEKLFQKI 106
+K++ Y + +Q + D N MF N YN G+ + AE ++++F +K+
Sbjct: 441 VKEQKRTEYLYLSQ-FVNDVNQMFENAKTYNPKGNAVFKCAETMQEVFDKKL 491

>C34C6.3 CE43092 WBGene00007916 EGF receptor\ /notch-like
protein#status:Partially_confirmed#UniProt:Q18424#protei
n_id:CAA91258.3
Length = 529

Score = 29.3 bits (64), Expect = 0.40, Method: Composition-based stats.
Identities = 12/33 (36%), Positives = 20/33 (60%)

Query: 55 IKKRLNNYYWNAQECIQDFNTMFTNCYIYNKP 87
I+KR +Y + Q C Q FN+ +C+ Y++P
Sbjct: 196 IEKRCFCSYGGFFGQRCDQKFNSQNDHCFAYDEP 228

Appendix B.3 Chromo-Domain BLAST Alignment

>K08H2.6 CE06164 WBGene00001995 locus:hpl-1 murine modifier 2
protein
like#status:Confirmed#UniProt:Q21370#protein_id:CAA9415
2.1
Length = 184

Score = 43.5 bits (101), Expect = 2e-05, Method: Composition-based stats.
Identities = 17/48 (35%), Positives = 29/48 (60%)

Query: 2 YAVVEKIIDRRVRKGVVEYLLKWKGYXXXXXXXXXXXXLDCQDLIQYE 49
+ VEK++++R+ +G EYY+KW+G+ L C +IQ+YE
Sbjct: 37 FVVEKVLNKRRLTRGGSEYYIKWQGFPESECSWEPIENLQCDRMIQEYE 84

>ZK1236.2 CE00380 WBGene00000414 locus:cec-
1#Nucleolin#status:Confirmed#UniProt:P34618#protein_id:
AAA28192.1
Length = 304

Score = 34.7 bits (78), Expect = 0.010, Method: Composition-based stats.
Identities = 14/25 (56%), Positives = 19/25 (76%)

Query: 2 YAVVEKIIDRRVRKGVVEYLLKWKGY 26
Y VE I++ R +KKG E+Y+KW GY
Sbjct: 8 YTVESILEHRKKKKGKSEFYIKWLG Y 32

>K01G5.2c CE25038 WBGene00001996 locus:hpl-2 'chromo' (CHRromatin
Organization MOfifier)
domain#status:Confirmed#UniProt:Q9U3C6#protein_id:CAB54

267.2
Length = 303

Score = 31.2 bits (69), Expect = 0.11, Method: Composition-based stats.
Identities = 13/49 (26%), Positives = 28/49 (57%), Gaps = 1/49 (2%)

Query: 2 YAVEKIIDRRVRK-GKVEYLLKWKGYXXXXXXXXXXXXLDCQDLIQQYE 49
+ VEK++D+R K G+ E+ ++W+G+ L C +++ ++E
Sbjct: 19 FMVEKVLDRKRTGKAGRDEFLLIQWQGFPESSWEPRENLCVEMLDEFE 67

>K01G5.2b CE25037 WBGene00001996 locus:hpl-2 'chromo' (CHRromatin
Organization MODifier)
domain#status:Confirmed#UniProt:017918#protein_id:CAB07
243.2
Length = 301

Score = 31.2 bits (69), Expect = 0.11, Method: Composition-based stats.
Identities = 13/49 (26%), Positives = 28/49 (57%), Gaps = 1/49 (2%)

Query: 2 YAVEKIIDRRVRK-GKVEYLLKWKGYXXXXXXXXXXXXLDCQDLIQQYE 49
+ VEK++D+R K G+ E+ ++W+G+ L C +++ ++E
Sbjct: 19 FMVEKVLDRKRTGKAGRDEFLLIQWQGFPESSWEPRENLCVEMLDEFE 67

>F32E10.2 CE04475 WBGene00017990 chromo domain of heterochromatin
protein#status:Confirmed#UniProt:Q19972#protein_id:AAA83
357.1
Length = 270

Score = 30.0 bits (66), Expect = 0.24, Method: Composition-based stats.
Identities = 13/26 (50%), Positives = 18/26 (69%)

Query: 1 EYAVEKIIDRRVRK GKVEYLLKWKGY 26
EYAVE+++ R KG Y ++WKGY
Sbjct: 86 EYAVERVLAHRKVKGSPLYLVQWKGY 111

>K01G5.2a CE16191 WBGene00001996 locus:hpl-2 'chromo' (CHRromatin
Organization MODifier)
domain#status:Confirmed#UniProt:017916#protein_id:CAB07
241.1
Length = 175

Score = 28.9 bits (63), Expect = 0.53, Method: Composition-based stats.
Identities = 13/49 (26%), Positives = 28/49 (57%), Gaps = 1/49 (2%)

Query: 2 YAVEKIIDRRVRK-GKVEYLLKWKGYXXXXXXXXXXXXLDCQDLIQQYE 49
+ VEK++D+R K G+ E+ ++W+G+ L C +++ ++E
Sbjct: 19 FMVEKVLDRKRTGKAGRDEFLLIQWQGFPESSWEPRENLCVEMLDEFE 67

Appendix B.4 Plant-Homeo-domain BLAST Alignment

>T14G8.1 CE03657 WBGene00000482 locus:chd-3 helicase-DNA-binding
like
protein#status:Confirmed#UniProt:Q22516#protein_id:CAA91
810.1
Length = 1787

Score = 83.2 bits (204), Expect = 3e-17, Method: Composition-based stats.
Identities = 28/43 (65%), Positives = 35/43 (81%)

Query: 1 FCRVCKDGGELLCCDTC PSSYHIHCLNPPLPEIPNGEWLCPRC 43

+CR+CK+ +L CDTCPSSYH +C++PPL EIP GEW CPRC
Sbjct: 330 YCRICKETSNILLCDTCPSSYHAYCIDPPLTEIPEGEWSCPRC 372

Score = 59.7 bits (143), Expect = 3e-10, Method: Composition-based stats.
Identities = 19/42 (45%), Positives = 27/42 (64%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGEWLCPRC 43
C VC GEL+ CDTC +YH+ C++ + + P G+W CP C
Sbjct: 268 CEVCNQDGEMLLCDTCTRAYHVACIDENMEQPPEGDWSCPHC 309

>F26F12.7 CE17716 WBGene00002637 locus:let-418 DNA
helicase#status:Confirmed#UniProt:Q19815#protein_id:AAC2
5894.1
Length = 1829

Score = 65.9 bits (159), Expect = 4e-12, Method: Composition-based stats.
Identities = 23/44 (52%), Positives = 32/44 (72%), Gaps = 1/44 (2%)

Query: 1 FCRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGE-WLCPRC 43
FC++CK+ LL CD+C S+H +C++PPL E+P E W CPRC
Sbjct: 319 FCKICKETENLLLCDSCVCSFHAYCIDPPLTEVPKEETWSCPRC 362

Score = 60.5 bits (145), Expect = 2e-10, Method: Composition-based stats.
Identities = 21/43 (48%), Positives = 27/43 (62%)

Query: 1 FCRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGE-WLCPRC 43
+C CK GELL CDTCP +YH C++ + E P G+W C C
Sbjct: 258 YCEECKQDGEMLLCDTCPRAYHTVCIDENMEEPPEGDWSCAHC 300

>ZK783.4 CE34152 WBGene00001470 locus:flt-
1#status:Partially_confirmed#UniProt:Q23590#protein_id:AA
C24421.2
Length = 1376

Score = 57.0 bits (136), Expect = 2e-09, Method: Composition-based stats.
Identities = 20/45 (44%), Positives = 28/45 (62%), Gaps = 2/45 (4%)

Query: 1 FCRVCK--DGGELLCCDTCPSYHIHCLNPPLPEIPNGEWLCPRC 43
C++CK DG E+L CD C S H+ C P + ++P G+W C RC
Sbjct: 1088 LCQICKSMDGDEMLVCDGCEGCHMECFRPRMTKVPEGDWFCQRC 1132

>C44B9.4 CE30897 WBGene00008081 locus:athp-1 S.pombe hypothetical
protein C27F7.07C
like#status:Partially_confirmed#UniProt:Q18605#protein_i
d:CAA97781.2
Length = 1150

Score = 50.4 bits (119), Expect = 2e-07, Method: Composition-based stats.
Identities = 16/42 (38%), Positives = 27/42 (64%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGEWLCPRC 43
C +C GG +LCC+ CP+S+H+ C+ ++P+ + C RC
Sbjct: 62 CGICSSGGNILCCEQCPASFHLACIGYESSDLPDDNFYCNRC 103

Score = 33.1 bits (74), Expect = 0.029, Method: Composition-based stats.
Identities = 15/42 (35%), Positives = 21/42 (50%), Gaps = 1/42 (2%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEI-PNGEWLCPR 42

C + D +L CD C +H C+ PPL + W+CPR
Sbjct: 224 CNLKDDWTRMLKCFCDLIWHQKCVTPPLIHVRAYFYWMCP R 265

>T12D8.1 CE42503 WBGene00011729 locus:set-16 PHD-finger. (2 domains), SET domain#status:Partially_confirmed#UniProt:046025#protein_id:CAB05024.2 Length = 2519

Score = 50.1 bits (118), Expect = 2e-07, Method: Composition-based stats. Identities = 22/46 (47%), Positives = 27/46 (58%), Gaps = 3/46 (6%)

Query: 2 CRVCKDGGE---LLCCDTCPSYHIHCLNPPLPEIPNGEWLCPRC 44
C C GG+ LL CD C SYHI+C+ P L +IP G W C C+

Sbjct: 524 CEGCGTGGDEANLLLCDECDVSYHIYCMKPLLDKIPQGPWRQCWCS 569

>ZK593.4 CE35704 WBGene00004319 locus:rbr-2 Human XE169 like#status:Partially_confirmed#UniProt:Q23541#protein_id:CAA93426.2 Length = 1477

Score = 49.3 bits (116), Expect = 4e-07, Method: Composition-based stats. Identities = 21/48 (43%), Positives = 28/48 (58%), Gaps = 5/48 (10%)

Query: 1 FCRVCKDGGE---LLCCDT--CPSSYHIHCLNPPLPEIPNGEWLCPRC 43
FC C +G + LL CD C + H +C +P L E+P GEW CP+C

Sbjct: 321 FCVACNEGKDEDLALLLCDIDGCNNGRHTYCCDPVLDEVPEGEWRCPKC 368

Score = 33.5 bits (75), Expect = 0.022, Method: Composition-based stats. Identities = 15/39 (38%), Positives = 21/39 (53%), Gaps = 2/39 (5%)

Query: 7 DGGELLCCDTCPSYHIHCL--NPPLPEIPNGEWLCPRC 43
D L C C S +H+ C +P L ++P G +LC RC

Sbjct: 1216 DSESTLTCIMCDSEFHVRCEWSPFLEKLPPEGCFLCVRC 1254

>F17A2.3 CE05646 WBGene00008902 PHD-finger.#status:Predicted#UniProt:Q19511#protein_id:CAA92158.1 Length = 463

Score = 45.4 bits (106), Expect = 6e-06, Method: Composition-based stats. Identities = 18/43 (41%), Positives = 27/43 (62%), Gaps = 1/43 (2%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLN-PPLPEIPNGEWLCPRC 43
C +C DGG ++ C+TCP+S+H CL +PE ++C RC

Sbjct: 30 CGMCADGGTIIWCETCPASFHAFCLGLKTIPEPEKDTFICHR C 72

>Y59A8A.2 CE44093 WBGene00013339 status:Partially_confirmed UniProt:Q9GRZ5#protein_id:CAC14404.2 Length = 599

Score = 40.4 bits (93), Expect = 2e-04, Method: Composition-based stats. Identities = 19/45 (42%), Positives = 24/45 (53%), Gaps = 3/45 (6%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEIP---NGEWLCPRC 43
CR + + CD C SYHI CL+PPL +P N W+C C

Sbjct: 518 CRKSTEQHKQTQCDECHKSYHIGCLSPLTRLPKRNNFGWICHEC 562

>K09A11.5 CE34205 WBGene00010708 PHD-finger.#status:Partially_confirmed#UniProt:Q21375#protein_id:CAA90618.2
Length = 650

Score = 40.0 bits (92), Expect = 2e-04, Method: Composition-based stats.
Identities = 19/43 (44%), Positives = 25/43 (58%), Gaps = 1/43 (2%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGE-WLCPRC 43
C +C GE+L C +CP+S+HI CL PNG + C RC
Sbjct: 53 CCICARRGEVLWCHSCPASFHIKCLGYDTPQNGTIFTCRRRC 95

>C28H8.9a CE06896 WBGene00016200 status:Confirmed UniProt:Q09477 protein_id:AAA62297.3
Length = 372

Score = 38.9 bits (89), Expect = 5e-04, Method: Composition-based stats.
Identities = 16/42 (38%), Positives = 24/42 (57%)

Query: 2 CRVCKDGGELLCCDTCPSYHIHCLNPPLPEIPNGEWLCPRC 43
C ++ +LL CD C YH++CL P L + P+ E+ C C
Sbjct: 317 CGTSENDKLLFCDDCDRGYHLYCLTPALEKAPDDEYSCRLC 358

>F33E11.6b CE39929 WBGene00018013 status:Confirmed UniProt:Q2A950 protein_id:ABD63225.1
Length = 447

Score = 36.2 bits (82), Expect = 0.003, Method: Composition-based stats.
Identities = 18/46 (39%), Positives = 23/46 (50%), Gaps = 4/46 (8%)

Query: 2 CRVC-KDGGELLCCDTCPSYHIHCLNPP---LPEIPNGEWLCPRC 43
C C K GGE++CC TC +YH C+ P + EW C C
Sbjct: 335 CDSCEKTGGEMICCATCKIAYHPQCIEMPERMAALVKTYEWSVDC 380

Score = 32.3 bits (72), Expect = 0.052, Method: Composition-based stats.
Identities = 14/45 (31%), Positives = 24/45 (53%), Gaps = 8/45 (17%)

Query: 2 CRVC-----KDGGELLCCDTCPSYHIHCLNPPLPEIPNGEWL C 40
CR+C + E++ CD C +H +C+ L ++P G W+C
Sbjct: 380 CRLCSICNKPEKEDEIVFCDRCDRGFHTYCVG--LKKLPQGTWIC 422

>H05L14.2 CE42798 WBGene00010367 Zinc finger, C3HC4 type (RING finger)#status:Partially_confirmed#UniProt:O17709#protein_id:CAB16922.3
Length = 2199

Score = 34.7 bits (78), Expect = 0.010, Method: Composition-based stats.
Identities = 16/52 (30%), Positives = 24/52 (46%), Gaps = 18/52 (34%)

Query: 2 CRVC---KDGGELLCCDTCPSYHIHCLNPPLPEIPNGEWL----CPRCT 44
C +C ++ E + CDTC YH HC++ WL CP+C+
Sbjct: 2144 CLICTEIIIEEAVETVTCDTCTREYHYHCIS-----RWLKINSVCPQCS 2186

>F26H11.2i CE43186 WBGene00009180 locus:nurf-1#status:Partially_confirmed#UniProt:B6VQ92#protein_id:CAR97823.1
Length = 1619

Score = 33.5 bits (75), Expect = 0.023, Method: Composition-based stats.
Identities = 13/34 (38%), Positives = 21/34 (61%), Gaps = 2/34 (5%)

Query: 2 CRVC-KDGGELLCCDTCPSSEYHIHCLN-PPLPEI 33
CRVC K G ++ C C +++H+ C + P PE+
Sbjct: 350 CRVCGKSSGRVVGCTQCEAAFHVVECSHLKPFPEV 383

>F26H11.2b CE35295 WBGene00009180 locus:nurf-
1#status:Partially_confirmed#UniProt:Q6BER5#protein_id:C
AC42289.2
Length = 1693

Score = 33.5 bits (75), Expect = 0.023, Method: Composition-based stats.
Identities = 13/34 (38%), Positives = 21/34 (61%), Gaps = 2/34 (5%)

Query: 2 CRVC-KDGGELLCCDTCPSSEYHIHCLN-PPLPEI 33
CRVC K G ++ C C +++H+ C + P PE+
Sbjct: 422 CRVCGKSSGRVVGCTQCEAAFHVVECSHLKPFPEV 455

>F26H11.2a CE35294 WBGene00009180 locus:nurf-
1#status:Partially_confirmed#UniProt:Q6BER5#protein_id:C
AB04197.2
Length = 1691

Score = 33.5 bits (75), Expect = 0.023, Method: Composition-based stats.
Identities = 13/34 (38%), Positives = 21/34 (61%), Gaps = 2/34 (5%)

Query: 2 CRVC-KDGGELLCCDTCPSSEYHIHCLN-PPLPEI 33
CRVC K G ++ C C +++H+ C + P PE+
Sbjct: 422 CRVCGKSSGRVVGCTQCEAAFHVVECSHLKPFPEV 455

>F26H11.2c CE36931 WBGene00009180 locus:nurf-
1#status:Partially_confirmed#UniProt:Q6BER5#protein_id:C
AH04722.1
Length = 2266

Score = 33.1 bits (74), Expect = 0.027, Method: Composition-based stats.
Identities = 13/34 (38%), Positives = 21/34 (61%), Gaps = 2/34 (5%)

Query: 2 CRVC-KDGGELLCCDTCPSSEYHIHCLN-PPLPEI 33
CRVC K G ++ C C +++H+ C + P PE+
Sbjct: 422 CRVCGKSSGRVVGCTQCEAAFHVVECSHLKPFPEV 455

>H20J04.2 CE27187 WBGene00019217 status:Partially_confirmed UniProt:Q9
N5L9#protein_id:AAF39888.2
Length = 1427

Score = 32.3 bits (72), Expect = 0.049, Method: Composition-based stats.
Identities = 12/45 (26%), Positives = 20/45 (44%), Gaps = 3/45 (6%)

Query: 2 CRVC-KDGG---ELLCCDTCPSSEYHIHCLNPPLPEIPNGEWLCPRC 43
CR C+ +L+ C C + YH+ C + +W+C C
Sbjct: 1115 CRSCRRKAAAHDLVLCSECDNCHLKAALDVNSDAPADWMCTSC 1159

>F42A9.2 CE07224 WBGene00003034 locus:lin-49 zinc-finger
protein#status:Confirmed#UniProt:Q20318#protein_id:AAB03
164.1
Length = 1042

Score = 32.3 bits (72), Expect = 0.051, Method: Composition-based stats.
Identities = 17/47 (36%), Positives = 22/47 (46%), Gaps = 7/47 (14%)

Query: 2 CRVCKDGDG----GELLCCDTCPSSEYHIHCLNPPLPEIPNGEWLCPRC 43
C +C DG +++ CD C S H C +P IP G C RC
Sbjct: 198 CNICLDGDTSNQIVYCDRCNLSVHQDCYG--IPFIPEGCLECRRC 242

>C11G6.3 CE05257 WBGene00007524 PHD-
finger.#status:Partially_confirmed#UniProt:Q17909#protei
n_id:CAA94113.1
Length = 385

Score = 31.6 bits (70), Expect = 0.083, Method: Composition-based stats.
Identities = 16/47 (34%), Positives = 22/47 (46%), Gaps = 5/47 (10%)

Query: 2 CRVCKD---GGELLCCDTCPSSEYHIHCLNPPLPEIPNGEWLCPRCT 44
C VC G ++ CD C +H HC+ E + +W C RCT
Sbjct: 310 CPVCSVAYTVGANMIGDQCQDWFHWHCVGLT-AEPTDSKWFCRTRCT 355

>F54F2.2a CE25003 WBGene00006975 locus:zfp-
1#status:Confirmed#UniProt:P34447#protein_id:AAK26137.1
Length = 867

Score = 30.4 bits (67), Expect = 0.20, Method: Composition-based stats.
Identities = 18/50 (36%), Positives = 23/50 (46%), Gaps = 9/50 (18%)

Query: 2 CRVCKD-----GGELLCCD--TCPSSEYHIHCLNPPLPEIPNGEWLCPRCT 44
C VC D L+ CD C + H C + E+P GEW C +CT
Sbjct: 8 CCVCADENGWTDNPLIYCDGENCEVAHVQGCY--IQEVPEGEWFCAKCT 55

>Y51H1A.4 CE20286 WBGene00013095 locus:ing-3 PHD-
finger.#status:Confirmed#UniProt:Q9XWJ8#protein_id:CAA21
665.1
Length = 490

Score = 29.6 bits (65), Expect = 0.28, Method: Composition-based stats.
Identities = 16/46 (34%), Positives = 24/46 (52%), Gaps = 6/46 (13%)

Query: 1 FCRVCKDGGELLCCDTCPS---YHIHCLNPPLPEIPNGEWLCPRC 43
FC K G+++ CD + +H C+ + E P G+W CPRC
Sbjct: 432 FCNE-KSYGDMVQCDNRHCTLRWFHYPCIG--MVEPPTGKWYCPRC 474

Appendix B.4 SET-domain BLAST Alignment

>C43E11.3b CE08681 WBGene00016603 locus:met-
1#status:Partially_confirmed#UniProt:A4LBC3#protein_id:A
B052817.1
Length = 1590

Score = 90.9 bits (224), Expect = 2e-19, Method: Composition-based stats.
Identities = 53/137 (38%), Positives = 75/137 (54%), Gaps = 9/137 (6%)

Query: 2387 GLGLYAKVDISMGDFIIEYKGEIRSEVCEVREIRYVAQNRGVYMFRIIDEE-WVIDATMA 2445
G GL A DI G FIIIEY GE++ + E R+ +Y A + + + D + IDAT+
Sbjct: 681 GCGLRAVKDIKGRFIIIEYIGEVEVREIRYVAQNRGVYMFRIIDEE-WVIDATMA 740

Query: 2446 GGPARYI~~NHSCDPN~~CSTQILDAGSGARE----KKIITANRPI~~SANEELTYDY~~QFELEGT 2501
G P+R++NHSCDPN I + S R ++ + R I A EE+T+DYQF G
Sbjct: 741 GNPSRFVNHSCDPNA---ICEKWSVPRTPGDVNRVGFVSKRFIKAGEEITFDYQFVNYG- 796

Query: 2502 TDKIPCLCGAPNCVKWM 2518
D C CG+ +C W+
Sbjct: 797 RDAQQCFCGSASCSGWI 813

>C43E11.3a CE30503 WBGene00016603 locus:met-
1#status:Partially_confirmed#UniProt:A4LBC2#protein_id:A
B052816.1
Length = 1604

Score = 90.9 bits (224), Expect = 2e-19, Method: Composition-based stats.
Identities = 53/137 (38%), Positives = 75/137 (54%), Gaps = 9/137 (6%)

Query: 2387 GLGLYAKVDISMGDFIIEYKGEIIRSEVCEVREIRYVAQNRGVYMFRIIDEE-WVIDATMA 2445
G GL A DI G FIIIEY GE++ + E R+ +Y A + + + D + IDAT+
Sbjct: 695 GCGLRAVKDIKGRFIIIEYIGEVVERDDYEKRKTKYAADKHKHHYLCDTGVYTIDATVY 754

Query: 2446 GGPARYINHSCDPNCSTQILDAGSGARE----KKIITANRPIISANEELTYDYQFELEGT 2501
G P+R++NHSCDPN I + S R ++ + R I A EE+T+DYQF G
Sbjct: 755 GNPSRFVNHSCDPNA--ICEKWSVPRTPGDVNRVGVFFSKRFKAGEEITFDYQFVNYG- 810

Query: 2502 TDKIPCLCGAPNCVKWM 2518
D C CG+ +C W+
Sbjct: 811 RDAQQCFCGSASCSGWI 827

>Y2H9A.1 CE27781 WBGene00003222 locus:mes-4 SET
domain#status:Confirmed#UniProt:Q9NH52#protein_id:CAA162
76.2
Length = 898

Score = 65.5 bits (158), Expect = 8e-12, Method: Composition-based stats.
Identities = 46/143 (32%), Positives = 70/143 (48%), Gaps = 15/143 (10%)

Query: 2376 DRVYLARSRIAGLGLYAKVDISMGDFIIEYKGEIIRSEVCEVREIRYVAQNRGV----YM 2431
+++ LA + G G++AK I ++I EY GEII + + R + V+ +R YM
Sbjct: 537 EKIKLAATLCKGYGVFAKQIEKDEYICEYVGEII-DKAEKRRRLDSVVISRDFQANHYM 595

Query: 2432 FRIDEEWVIDATMAGGPARYINHSCDPNCSTQILDA-----GSGAREKKIITANRPIIS 2485
+ + +DA G +RYINHSCDPN ++ + + + I A R I
Sbjct: 596 MELHKGLTVDAARYGNISRYINHSCDPNAASFVTKVFKKTEKESLYDTRSYIRAIRTID 655

Query: 2486 ANEELTYDYQFELEGTDDKIP-CLCGAPNCVKWM 2518
+E+T+ Y E + +P C CGA NC+ M
Sbjct: 656 DGDEITFSYMNNE---ENLPDCECGAENCMGTM 686

>R06A4.7 CE28067 WBGene00003220 locus:mes-2 SET
domain#status:Confirmed#UniProt:O17514#protein_id:CAB0558
9.2
Length = 773

Score = 60.5 bits (145), Expect = 1e-08, Method: Composition-based stats.
Identities = 45/131 (34%), Positives = 64/131 (48%), Gaps = 5/131 (3%)

Query: 2370 MRREWKDRVYLARSRIAGLGLYAKVDISMGDFIIEYKGEIIRSEVCEVREIRYVAQNRGV 2429
M R + R Y S+IAG GL+ +FI EY GE I + E R Y + +
Sbjct: 615 MTRMIQKRTYCGPSKIAGNGLFLLPEAEKDEFITEYTGERISDDEAERRGAIY-DRYQCS 673

Query: 2430 YMFRIIDEEWVIDATMAGGPARYINH-SCDPNCSTQILDAGSGAREKKIITANRPIISANE 2488
Y+F I+ ID+ G AR+ NH S +P C + + A E +I A R + +E
Sbjct: 674 YIFNIETGGAIDSYKIGNLARFANHDSKNPTCYARTMVV---AGEHRIGFYAKRRLEISE 730

Query: 2489 ELTYDYQFELE 2499
ELT+DY + E
Sbjct: 731 ELTFDYSYSGE 741

>[R05D3.11](#) CE42016 WBGene00019883 locus:met-
2#status:Partially_confirmed#UniProt:P34544#protein_id:AA
K21437.2
Length = 1316

Score = 59.7 bits (143), Expect = 5e-10, Method: Composition-based stats.
Identities = 30/79 (37%), Positives = 44/79 (55%), Gaps = 2/79 (2%)

Query: 2438 WVIDATMAGGPARYINHSCDPNCSTQ-ILDAGSGAREKKIIITANRPISANEELTYDYQF 2496
+VIDA G R++NHSCDPN Q ++ R + + + A +ELT+DYQ+
Sbjct: 1231 YVIDAKQRGNLGRFLNHSCDPNVHVQHVMYDTHDLRLPWVAFTRKYVVKAGDELTDYQY 1290

Query: 2497 ELEGT-TDKIPCLCGAPNC 2514
+ T T ++ C CGA NC
Sbjct: 1291 TQDQTATTQLTCHCGAENC 1309

>[T12F5.4](#) CE13601 WBGene00003040 locus:lin-
59#status:Confirmed#UniProt:044757#protein_id:AAB96746.1
Length = 1312

Score = 52.8 bits (125), Expect = 6e-08, Method: Composition-based stats.
Identities = 40/138 (28%), Positives = 58/138 (42%), Gaps = 24/138 (17%)

Query: 2367 YQKMRREWKD----RVYLARSRIAGLGLYAKVDISMGDFIIEYKGEIRSEVCEVREIRY 2422
Y RR WK+ ++ ++ + L K+ G+F+ EY GE+I E + +
Sbjct: 625 YCSNRRFWKEDCGNKLCVSNNGPRSKRVLTKIARRAGEFLCEYAGEVITREQAQEK---- 680

Query: 2423 VAQNRGVYMFRIDEEWVIDATMAGGPARYINHSCDPNCSTQILDAGSGAREKKIIITANR 2482
AQ+R + I +DAT AR+I HSC PN ++ R ++
Sbjct: 681 FAQDRPRIIAIAAHLFVDATKRSNIARFIKHSCKPNSRLEVWSVNGFYRAGVFALSDLN 740

Query: 2483 PISANEELTYDYQFELEGTDDKIP----CLCGAPNC 2514
P N E+T D +D +P C CGA C
Sbjct: 741 P---NAEITVD-----KDLLPFDMACNCGATEC 766

Appendix B.4 SET-domain BLAST Alignment (JHDM1)

>[T26A5.5a](#) CE32733 WBGene00020821 status:Partially_confirmed UniProt:
Q95Q98#protein_id:AAN65291.1
Length = 1076

Score = 385 bits (988), Expect = e-108, Method: Composition-based stats.
Identities = 173/173 (100%), Positives = 173/173 (100%)

Query: 1 FSQTPLEDLVKSPPELVLRQIDWVGNGQWPDALRQRWISFNDRDCKKFFYNPHHTFPKVQNYCLM 60
FSQTPLEDLVKSPPELVLRQIDWVGNGQWPDALRQRWISFNDRDCKKFFYNPHHTFPKVQNYCLM
Sbjct: 93 FSQTPLEDLVKSPPELVLRQIDWVGNGQWPDALRQRWISFNDRDCKKFFYNPHHTFPKVQNYCLM 152

Query: 61 SVANCYTDFHIDFSGTSSVWYHVLKGRKVFVWLIPPTETNFFIYQEFIKTVNDNAFFGKSVE 120
SVANCYTDFHIDFSGTSSVWYHVLKGRKVFVWLIPPTETNFFIYQEFIKTVNDNAFFGKSVE
Sbjct: 153 SVANCYTDFHIDFSGTSSVWYHVLKGRKVFVWLIPPTETNFFIYQEFIKTVNDNAFFGKSVE 212

Query: 121 KCHVAILEPGDTMLIPSGWIHAVYTPDDSLVFGGNFLHSQSCKTQLRVYQVEN 173
KCHVAILEPGDTMLIPSGWIHAVYTPDDSLVFGGNFLHSQSCKTQLRVYQVEN
Sbjct: 213 KCHVAILEPGDTMLIPSGWIHAVYTPDDSLVFGGNFLHSQSCKTQLRVYQVEN 265

>[T26A5.5b](#) CE32734 WBGene00020821 status:Confirmed UniProt:Q95Q98 protein_id: AAN65292.1 Length = 505

Score = 381 bits (978), Expect = e-106, Method: Composition-based stats. Identities = 173/173 (100%), Positives = 173/173 (100%)

Query: 1 FSQTPLEDLVKSPPELVRQIDWVGNGQWPDALRQRWISFNDRDCKFYNPHTFPKVQNYCLM 60
Sbjct: 93 FSQTPLEDLVKSPPELVRQIDWVGNGQWPDALRQRWISFNDRDCKFYNPHTFPKVQNYCLM 152

Query: 61 SVANCYTDFHIDFSGTSVWYHVLKGRKVFVWLIPTTETNFFIYQEFIKTVNDNAFFGKSVE 120
Sbjct: 153 SVANCYTDFHIDFSGTSVWYHVLKGRKVFVWLIPTTETNFFIYQEFIKTVNDNAFFGKSVE 212

Query: 121 KCHVAILEPGDTMLIPSGWIHAVYTPDSSLVFGGNFLHSQSCKTQLRVYQVEN 173
Sbjct: 213 KCHVAILEPGDTMLIPSGWIHAVYTPDSSLVFGGNFLHSQSCKTQLRVYQVEN 265

>[F29B9.2a](#) CE09781 WBGene00017920 status:Confirmed UniProt:Q9GYI0 protein_id: AAK29799.1 Length = 910

Score = 157 bits (398), Expect = 2e-39, Method: Composition-based stats. Identities = 74/169 (43%), Positives = 105/169 (62%), Gaps = 3/169 (1%)

Query: 6 LEDLVKSPPELVRQIDWVGNGQWPDALRQRWISFNDRDCKFYNPHTFPKVQNYCLMSVANC 65
Sbjct: 446 MKEIAKPPRFVQEISMVNLWPDVSGAEYIKLLQREE--YLPEDQRPKVEQFCLAGMAGS 503

Query: 66 YTDFHIDFSGTSVWYHVLKGRKVFVWLIPTTETNFFIYQEFIKTVNDNAFFGKSVE-KCHV 124
Sbjct: 504 YTDFHVDVDFGGSSVYHILKGEKIFFYAAPEQNFQAAHETSPTTTWFGDIANGAVKR 563

Query: 125 AILEPGDTMLIPSGWIHAVYTPDSSLVFGGNFLHSQSCKTQLRVYQVEN 173
Sbjct: 564 VVIKEGQTLIPAGWIHAVLTPVDSLFGGNFLHNLGNLQMQRVYHLEN 612

>[F29B9.2b](#) CE27145 WBGene00017920 status:Confirmed UniProt:Q9BI67 protein_id: AAK29800.1 Length = 897

Score = 157 bits (397), Expect = 2e-39, Method: Composition-based stats. Identities = 74/169 (43%), Positives = 105/169 (62%), Gaps = 3/169 (1%)

Query: 6 LEDLVKSPPELVRQIDWVGNGQWPDALRQRWISFNDRDCKFYNPHTFPKVQNYCLMSVANC 65
Sbjct: 433 MKEIAKPPRFVQEISMVNLWPDVSGAEYIKLLQREE--YLPEDQRPKVEQFCLAGMAGS 490

Query: 66 YTDFHIDFSGTSVWYHVLKGRKVFVWLIPTTETNFFIYQEFIKTVNDNAFFGKSVE-KCHV 124
Sbjct: 491 YTDFHVDVDFGGSSVYHILKGEKIFFYAAPEQNFQAAHETSPTTTWFGDIANGAVKR 550

Query: 125 AILEPGDTMLIPSGWIHAVYTPDSSLVFGGNFLHSQSCKTQLRVYQVEN 173
Sbjct: 551 VVIKEGQTLIPAGWIHAVLTPVDSLFGGNFLHNLGNLQMQRVYHLEN 599

>[F43G6.6](#) CE20788 WBGene00005013 status:Partially_confirmed UniProt:Q20367#protein_id:CAA90395.1 Length = 548

Score = 132 bits (333), Expect = 6e-32, Method: Composition-based stats.
Identities = 67/175 (38%), Positives = 101/175 (57%), Gaps = 13/175 (7%)

Query: 1 FSQTP-LEDLVKSPQLVLRQIDWVGNQWPDALRQRWISFNDRDCKKFYNPHHTFPKQVNYCL 59
FS P L+++ + P V+ I W D + +S + R PK++ C
Sbjct: 223 FSDHPELKEMARPPRFVQDISMAKRLWSDVTSKALSDDHR-----PKIEQICA 271

Query: 60 MSVANCY^TDF^HI^DFSGTSVWYHVLKGR^KVFWLIPPTETNFFIYQEFIKTVNDNAFFGKSV 119
++AN YTDFH+DF GTSV++HV KG K+F++ PTE NF +YQ + + + + G ++
Sbjct: 272 AAMANSYTDHFVDFGGTSVYFHVFKGEKIFUYIAAPTEENFVMYQAHETSTDSSIWLGHTL 331

Query: 120 EKC-HVAILEPGDMLIPSGWI^HAVYTPDDSLVFGGNFLHSQSCKTQLRVYQVEN 173
+ +++ G T+LIP+GWIHAV T DSL FGGNFLH + +RV +EN
Sbjct: 332 KGALKRVVVKEGQTLIPAGWIHAVLTTIDSLAFGGNFLHLGNLIMHMRVVDMEN 386

>[F29B9.4a](#) CE27146 WBGene00004205 locus:psr-
1#status:Confirmed#UniProt:Q9GYI4#protein_id:AAF99922.2
Length = 400

Score = 44.3 bits (103), Expect = 4e-05, Method: Composition-based stats.
Identities = 40/135 (29%), Positives = 58/135 (42%), Gaps = 22/135 (16%)

Query: 37 FNGRDCKKFYNPHHTFPKQVNYCLMSVANCY^TDF^HI^DFSGTSVWYHVLKGR^KVFWLIPPT 96
F+ D K PH F +M A T HID GTS W +L+G K + LIPP
Sbjct: 166 FHYADDKRPPHRWF-----VMGPARGTAIHIDPLGTSAWNSLLQGHKRWLIPPIA 218

Query: 97 TNFFI---YQEFIKTVNDNAFFGKSVEK-----CHVAILE----PGDMLIPSGWI^H 141
+ E K ++ + ++V K A +E PG+TM +PSGW H
Sbjct: 219 PRDLVKPMAHEKGKHPDEGITWFQTVYKRVRSWPKEYAPIECRQGPGETMFVPSGWWH 278

Query: 142 AVYTPDDSLVFGGNF 156
V + ++ N+
Sbjct: 279 VVINEEYTIAVTHNY 293

>[T07C4.11](#) CE40266 WBGene00011563 status:Partially_confirmed UniProt:
Q14V35#protein_id:CAK55173.1
Length = 367

Score = 43.5 bits (101), Expect = 5e-05, Method: Composition-based stats.
Identities = 25/111 (22%), Positives = 52/111 (46%), Gaps = 13/111 (11%)

Query: 57 YCLMSVANCY^TDF^HI^DFSGTSVWYHVLKGR^KVFWLIPPTETNFFIYQEFIKTVNDNAF-- 114
+ + + +T H D + W + GRK +++++PP N F +V ++ F
Sbjct: 139 FVYIGASGSWTKLHSDVVSWSANICGRKQWFMPPGSENLF-----SSVTESGFVD 193

Query: 115 ----FGKSVEKCHVA--ILEPGDMLIPSGWI^HAVYTPDDSLVFGGNFLHS 159
+ + E+ V + EPG+ + +PS W H + +D++ N+++S
Sbjct: 194 DIREYERLFEQAKVIKVFQEPGEIVFVPSNWHYQAHNLEDTISINHNWMS 244

>[F29B9.4b](#) CE39926 WBGene00004205 locus:psr-
1#status:Confirmed#UniProt:Q27GT3#protein_id:ABD63227.1
Length = 284

Score = 43.5 bits (101), Expect = 5e-05, Method: Composition-based stats.
Identities = 40/135 (29%), Positives = 58/135 (42%), Gaps = 22/135 (16%)

Query: 37 FNGRDCKKFYNPHHTFPKQVNYCLMSVANCY^TDF^HI^DFSGTSVWYHVLKGR^KVFWLIPPT 96
F+ D K PH F +M A T HID GTS W +L+G K + LIPP

Sbjct: 50 FHYADDKRPPHRWF-----VMGPARSGTAIHIDPLGTSAWNSLLQGHKRWVLIPPIA 102
 Query: 97 TNFFI---YQEFIKTVNDNAFFGKSVEK-----CHVAILE----PGDTMLIPSGWIH 141
 + E K ++ + ++V K A +E PG+TM +PSGW H
 Sbjct: 103 PRDLVKPMAHEKGKHPDEGITWFQTVYKRVRSWPKEYAPIECRQGPGETMFVPSGWWH 162
 Query: 142 AVYTPDDSLVFGGNF 156
 V + ++ N+
 Sbjct: 163 VVINEEYTIAVTHNY 177
(JARID1/2)

>[ZK593.4](#) CE35704 WBGene00004319 locus:rbr-2 Human XE169
 like#status:Partially_confirmed#UniProt:Q23541#protein_id:CAA93426.2
 Length = 1477

Score = 241 bits (615), Expect = 5e-65, Method: Composition-based stats.
 Identities = 104/104 (100%), Positives = 104/104 (100%)

Query: 1 GMCFSTFCWHTEDHWTYSVNYNHFGGERKIWYGVGGEDA EK FEDALKKIAPGLTGRQRDLF 60
 GMCFSTFCWHTEDHWTYSVNYNHFGGERKIWYGVGGEDA EK FEDALKKIAPGLTGRQRDLF
 Sbjct: 505 GMCFSTFCWHTEDHWTYSVNYNHFGGERKIWYGVGGEDA EK FEDALKKIAPGLTGRQRDLF 564
 Query: 61 HHMTTAANPHLLRSLGVPIHSVHQNAGEFVITFPRAYHAGFNEG 104
 HHMTTAANPHLLRSLGVPIHSVHQNAGEFVITFPRAYHAGFNEG
 Sbjct: 565 HHMTTAANPHLLRSLGVPIHSVHQNAGEFVITFPRAYHAGFNEG 608

>[Y48B6A.11](#) CE41181 WBGene00012982 locus:jmjd-
 2#status:Partially_confirmed#UniProt:Q9U297#protein_id:CA54451.2
 Length = 922

Score = 86.7 bits (213), Expect = 2e-18, Method: Composition-based stats.
 Identities = 44/111 (39%), Positives = 59/111 (53%), Gaps = 11/111 (9%)

Query: 1 GMCFSTFCWHTEDHWTYSVNYNHFGGERKIWYGVGGEDA EK FEDALKK-----IAPGLT 53
 GM +TF WH ED YS+N+ HFG K W+ + E A++FE + + AP
 Sbjct: 256 GMYKTFPWAHEDMDLYSINFLHFGAPKYWFAISSEHADRFRERFMSQQFSYQNEYAP--- 312
 Query: 54 GRQRDLFHMTTAANPHLLRSLGVPIHSVHQNAGEFVITFPRAYHAGFNEG 104
 + + H T P LLR G+P ++ Q EF+ITFPR YH GFN G
 Sbjct: 313 -QCKAFLRHKTYLVTEPELLRQAGIPYATMVQRPNEFIITFPRGYHMGFNLG 362

>[C29F7.6](#) CE08447 WBGene00007813 status:Partially_confirmed UniProt:O17619#protein_id:CAB07325.1
 Length = 732

Score = 30.8 bits (68), Expect = 0.14, Method: Composition-based stats.
 Identities = 26/94 (27%), Positives = 39/94 (41%), Gaps = 11/94 (11%)

Query: 6 TFCWHTEDHWTYSVNYNHFGGERKIWYGVGGEDA EK FEDALKKIAPGLTGRQRDLFHMTT 65
 T C H E+ S+N N + IWY V E + KFE L K ++L+ + +
 Sbjct: 507 TTC-HIENQAIGSLNLLGPGKCIWYAVASEHSAKFEQLLMK-----KNLWPYDSV 556
 Query: 66 A-ANPHLLRSLGVPIHSVHQNAGEFVITFPRAYH 98
 N L + G+P+ Q + V YH
 Sbjct: 557 LWPNEEELLNWGI PVMKFIQETDDTVYVGTGTYH 590

>[C16C10.2](#) CE01493 WBGene00007623 status:Confirmed UniProt:Q09462 p
rotein_id:CAA86740.1
Length = 262

Score = 28.5 bits (62), Expect = 0.73, Method: Composition-based stats.
Identities = 12/37 (32%), Positives = 20/37 (54%)

Query: 26 ERKIWYGVGGEDA EK FEDALKKIAPGLTGRQRDLFHH 62
E+K Y + ED +K D +KK+ + +D +HH
Sbjct: 33 EKKKDYKLRAEDYQKKRDTIKKLLKKSAMDKNQDEYHH 69

>[F23D12.5](#) CE15893 WBGene00009089 status:Partially_confirmed UniProt:
Q19760#protein_id:CAA94916.1
Length = 867

Score = 28.1 bits (61), Expect = 0.91, Method: Composition-based stats.
Identities = 14/38 (36%), Positives = 20/38 (52%)

Query: 10 **HTE**DHWTSVNYNHFGERKIWYGVGGEDA EK FEDALKK 47
H E+ S+N NH +WYGV E + + E +KK
Sbjct: 624 HLENQALGSININHGPGDCVWYGVPM EYSGRMEVLIKK 661
(JHDM3/JMJD2)

>[C29F7.6](#) CE08447 WBGene00007813 status:Partially_confirmed UniProt:0
17619#protein_id:CAB07325.1
Length = 732

Score = 49.3 bits (116), Expect = 9e-07, Method: Composition-based stats.
Identities = 41/144 (28%), Positives = 67/144 (46%), Gaps = 23/144 (15%)

Query: 13 GTILEDTN YE IKG VNTVYLYFGMYKTT**FPWHA**EDMDLYSINFLHFGAP**K**-YWF A ISSEHA 71
G + E I GVN V +YF + H E+ + S+N L+ G K W+A++SEH+
Sbjct: 480 GGLNEYIKESIGGVNEVQMYFKQPGSRTTCHIENQAIGSLN-LNLGPGKCIWYAVASEHS 538

Query: 72 DRFERFMSQQ--FSYQNEYAPQCKAFLRHKTYLVTPPELLRQAGIPYATMVQRPNEFIITF 129
+FE+ + ++ + Y + P E L GIP +Q ++ +
Sbjct: 539 AKFEQLLMKKNLWPYDSVLWPN-----EEELLNWGI PVMKFIQETDDTVYVG 585

Query: 130 PRGY**H**----MGF--NLGYNLAEST 147
YH +GF N+ +N+AEST
Sbjct: 586 TGT YH WQSIGFTGNVSWNIAEST 609

>[F18E9.5b](#) CE30958 WBGene00017571 locus:tag-
279#status:Partially_confirmed#UniProt:Q95QK3#protein_id
:AAM54191.1
Length = 1061

Score = 40.4 bits (93), Expect = 5e-04, Method: Composition-based stats.
Identities = 31/127 (24%), Positives = 53/127 (41%), Gaps = 15/127 (11%)

Query: 10 NRLGTILEDTN YE IKG VNTVYLYFGMYKTT**FPWHA**EDMDLYSINFLHFGAP**K**YWF A ISSE 69
NR G +L ++ G+NTV +Y + P H E+ + SIN+ WFA+ E
Sbjct: 778 NREGNLLNYAGVDVLGINTVQMYAKPIGSRTPAHMENSLMASINWNRGPGTCVWFAVPYE 837

Query: 70 HADRFERFMSQQ-FSYQNE-YAPQCKAFLRHKTYLVTPPELLRQAGIPYATMVQRPNEFII 127
+ + E + + YQ++ Y P K L + G+P Q+ +E +
Sbjct: 838 YWQLEFMIGE HGHKYQDQDYWPSEKELL-----ELGVPVIKFEQKADEMVY 884

Query: 128 TFPRGYH 134
+H
Sbjct: 885 VNTGCFH 891

>[F18E9.5a](#) CE30957 WBGene00017571 locus:tag-
279#status:Partially_confirmed#UniProt:Q19565#protein_id
:AAM54190.1
Length = 1020

Score = 40.4 bits (93), Expect = 5e-04, Method: Composition-based stats.
Identities = 31/127 (24%), Positives = 53/127 (41%), Gaps = 15/127 (11%)

Query: 10 NRLGTILEDNTYEIKGVNTVYLYFGMYKTTFPWHAEDMDLYSINFLHFGAPKYWFAISSE 69
NR G +L ++ G+NTV +Y + P H E+ + SIN+ WFA+ E
Sbjct: 737 NREGNLLNYAGVDVLGINTVQMYAKPIGSRTPAHMENSMLASINWNRGPGTCVWFVAVPYE 796

Query: 70 HADRFERFMSQQ-FSYQNE-YAPQCKAFLRHKTYLVTPELLRQAGIPYATMVQRPNEFII 127
+ + E + + YQ++ Y P K L + G+P Q+ +E +
Sbjct: 797 YWGQLEFMIGEHEGHKYQDQDYWPSEKELL-----ELGVPVIKFEQKADEMVY 843

Query: 128 TFPRGYH 134
+H
Sbjct: 844 VNTGCFH 850

>[F23D12.5](#) CE15893 WBGene00009089 status:Partially_confirmed UniProt:
Q19760#protein_id:CAA94916.1
Length = 867

Score = 34.3 bits (77), Expect = 0.031, Method: Composition-based stats.
Identities = 30/143 (20%), Positives = 54/143 (37%), Gaps = 21/143 (14%)

Query: 13 GTILEDNTYEIKGVNTVYLYFGMYKTTFPWHAEDMDLYSINFLHFGAPKYWFAISSEHAD 72
G +L + G+N +Y H E+ L SIN H W+ + E++
Sbjct: 594 GNLLNFAQESLAGLNKPVYCKPPGARTTAHLENQALGSININHGPGDCVWYGVPMYVSG 653

Query: 73 RFERFMSQQF--SYQNEYAPQCKAFLRHKTYLVTPELLRQAGIPYATMVQRPNEFIITFP 130
R E + + Y++ Y P + + LR IP +Q+P + +
Sbjct: 654 RMEVLIKKHRLNVYKSGYWP-----SEQELRNEKIPSQKFLQKPGDMVYVGI 700

Query: 131 RGYH-----MGFNLGYNLAEST 147
+H N+ +N+A+ T
Sbjct: 701 GTFHWVQSNDFAINVSWNVAQPT 723

(UTX/UTY)

>D2021.1 CE01878 WBGene00017046 locus:utx-1 glucose repression
mediator
protein#status:Partially_confirmed#UniProt:Q09519#protein
_id:AAB36864.1
Length = 1168

Score = 375 bits (962), Expect = e-105, Method: Composition-based stats.
Identities = 164/164 (100%), Positives = 164/164 (100%)

Query: 1 KWGKQINELSKLPAFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMAS 60
KWGKQINELSKLPAFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMAS
Sbjct: 863 KWGKQINELSKLPAFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMAS 922

Query: 61 ININIGPGDCEWFAVPYEWGKMHKLCEKNGVDLLTGTFWPIIDDLLDAGIPVHRFTQKA 120
ININIGPGDCEWFAVPYEWGKMHKLCEKNGVDLLTGTFWPIIDDLLDAGIPVHRFTQKA

Sbjct: 923 ININIGPGDCEWFAVPYEWGKMHKLCEKNGVDLLTGTFWPIIDLLDAGIPVHRFTQKA 982
Query: 121 GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSYEY 164
GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSYEY
Sbjct: 983 GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSYEY 1026

>F18E9.5a CE30957 WBGene00017571 locus:tag-
279#status:Partially_confirmed#UniProt:Q19565#protein_id
:AAM54190.1
Length = 1020

Score = 169 bits (428), Expect = 6e-43, Method: Composition-based stats.
Identities = 75/161 (46%), Positives = 110/161 (68%)

Query: 4 KQINELSKLPFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMASINI 63
KQ+NE+ KLP F N+L++ G V G+NTV+++ K G RTPAH +++ MASIN
Sbjct: 722 KQMNEIEKLPFTLLPNREGNLLNYAGVDVLGINTVQMYAKPIGSRTPAHMENSLMASINW 781

Query: 64 NIGPGDCEWFAVPYEWGKMHKLCEKNGVDLLTGTFWPIIDLLDAGIPVHRFTQKAGDM 123
N GPG C WFAVPYEWG++ + ++G +WP +LL+ G+PV +F QKA +M
Sbjct: 782 NRGPGTCVWFAVPYEWGQLEFMIGEHHGKYQDQDYWPSEKELLELGPVVIKFEQKADEM 841

Query: 124 VYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSYEY 164
VYV+ G HWVQ++ +C N+SWNV NF QL+ S+++++
Sbjct: 842 VYVNTGCFHWVQNSFCINVSWNVGPNTQLATSIVAHDH 882

>F18E9.5b CE30958 WBGene00017571 locus:tag-
279#status:Partially_confirmed#UniProt:Q95QK3#protein_id
:AAM54191.1
Length = 1061

Score = 169 bits (428), Expect = 6e-43, Method: Composition-based stats.
Identities = 75/161 (46%), Positives = 110/161 (68%)

Query: 4 KQINELSKLPFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMASINI 63
KQ+NE+ KLP F N+L++ G V G+NTV+++ K G RTPAH +++ MASIN
Sbjct: 763 KQMNEIEKLPFTLLPNREGNLLNYAGVDVLGINTVQMYAKPIGSRTPAHMENSLMASINW 822

Query: 64 NIGPGDCEWFAVPYEWGKMHKLCEKNGVDLLTGTFWPIIDLLDAGIPVHRFTQKAGDM 123
N GPG C WFAVPYEWG++ + ++G +WP +LL+ G+PV +F QKA +M
Sbjct: 823 NRGPGTCVWFAVPYEWGQLEFMIGEHHGKYQDQDYWPSEKELLELGPVVIKFEQKADEM 882

Query: 124 VYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSYEY 164
VYV+ G HWVQ++ +C N+SWNV NF QL+ S+++++
Sbjct: 883 VYVNTGCFHWVQNSFCINVSWNVGPNTQLATSIVAHDH 923

>F23D12.5 CE15893 WBGene00009089 status:Partially_confirmed UniProt:
Q19760#protein_id:CAA94916.1
Length = 867

Score = 145 bits (365), Expect = 1e-35, Method: Composition-based stats.
Identities = 64/164 (39%), Positives = 102/164 (62%)

Query: 1 KWGKQINELSKLPFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMAS 60

++ +Q++E+ KLP R N+L+ + G+N +++ K PG RT AH ++ + S
 Sbjct: 573 RFKEQLDEIKKLPDCLRPDGAGNLLNFAQESLAGLNKPKQVYCKPPGARTTAHLENQALGS 632
 Query: 61 ININIGPGDCEWFAVPY EYWGKMHKLCEKNGVDLLTGTFWPIIDDLLDAGIPVHRFTQKA 120
 ININ GPGDC W+ VP EY G+M L +K+ +++ +WP +L + IP +F QK
 Sbjct: 633 ININHGPGDCVWYGVPMEYSGRMEVLIKKHRLNVYKSGYWPSEQELRNEKIPSQKFLQKP 692
 Query: 121 GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSY EY 164
 GDMVYV G HWVQ++ + N+SWNVA F QL+ +++ +++
 Sbjct: 693 GDMVYVGIGTFHWVQSNDFAINVSWNVAQPTFNQLAAAMVIHDH 736

>C29F7.6 CE08447 WBGene00007813 status:Partially_confirmed UniProt:0
 17619#protein_id:CAB07325.1
 Length = 732

Score = 132 bits (331), Expect = 1e-31, Method: Composition-based stats.
 Identities = 61/164 (37%), Positives = 95/164 (57%), Gaps = 2/164 (1%)

Query: 1 KWGKQINELSKLP AFCRLIAGSNMLSHLGHQVHGMNTVKLFMKVPGCRTPAHQDSNHMAS 60
 K+ I EL KLP F + G N ++ + G+N V+++ K PG RT H ++ + S
 Sbjct: 461 KFQPLIQELDKLPNFKTKGGLN--EYIKESIGGVNEVQMYFKQPGSRTTCHIENQAIGS 518
 Query: 61 ININIGPGDCEWFAVPY EYWGKMHKLCEKNGVDLLTGTFWPIIDDLLDAGIPVHRFTQKA 120
 +N+N+GPG C W+AV E+ K +L K + WP ++LL+ GIPV +F Q+
 Sbjct: 519 LNLNLGPGKCIWYAVASEHS AKFEQLLMKKNLWPYDSVLWPNEEELLNWGIPVMKFIQET 578
 Query: 121 GDMVYVSGGAIHWVQASGWCNNISWNVAPLNFQQLSISLLSY EY 164
 D VYV G HWVQ+ G+ N+SWN+A F Q +++ L +++
 Sbjct: 579 DDTVYVGTGTYHWVQSIGFTGNVSWNIAESTFDQFAMAALVHDH 622